

-Passaic River Study Area-

Data Presentation

September 26, 2002
Newark, New Jersey



Agenda

- 8:30 a.m.
Breakfast at NJTPA
(receipts available)
- 9:00 a.m.
Introduction
(G. Mancini)
 - Participant introductions
 - Safety/logistics/housekeeping
 - Orientation and presentation format
- 9:15 a.m.
PCB Sources Identification
(including a brief dioxin sources identification preview)
(D. Farley)
- 10:15 a.m.
Habitat Quantification
(D. Ludwig)
 - Summary and Interpretation
- 10:45 a.m. BREAK
- 11:00 a.m.
Benthic Community Analysis
(T. Iannuzzi)
 - Summary and interpretation
- 11:30 a.m.
Fish Community Analysis
(D. Ludwig)
 - Summary and interpretation
- 12:00 a.m. LUNCH
- 1:30 p.m.
Preliminary Sediment Quality
Triad (SQT) & Toxicity Identifica-
tion Evaluation (TIE) Analysis
(T. Iannuzzi)
 - Summaries and interpretation
- 2:30 p.m.
Topical Discussions and Q&A
(All)

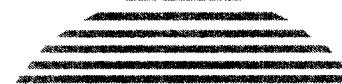




Agenda (cont'd)

- 3:00 p.m. Action Items/Next Meeting (G. Mancini)
 - Possible dates
 - Dioxin sources identification analyses
 - Other presentations?
- 5:00 p.m. ADJOURN

3



Meeting Overview

- Welcome and participant introductions
- Safety, logistics and housekeeping
- Handouts and supplemental materials
- Agenda and format

4





Meeting Objectives

- Summarize and interpret data
- Characterize study area
- Present and discuss source analyses
- Engage Q&A and discussion

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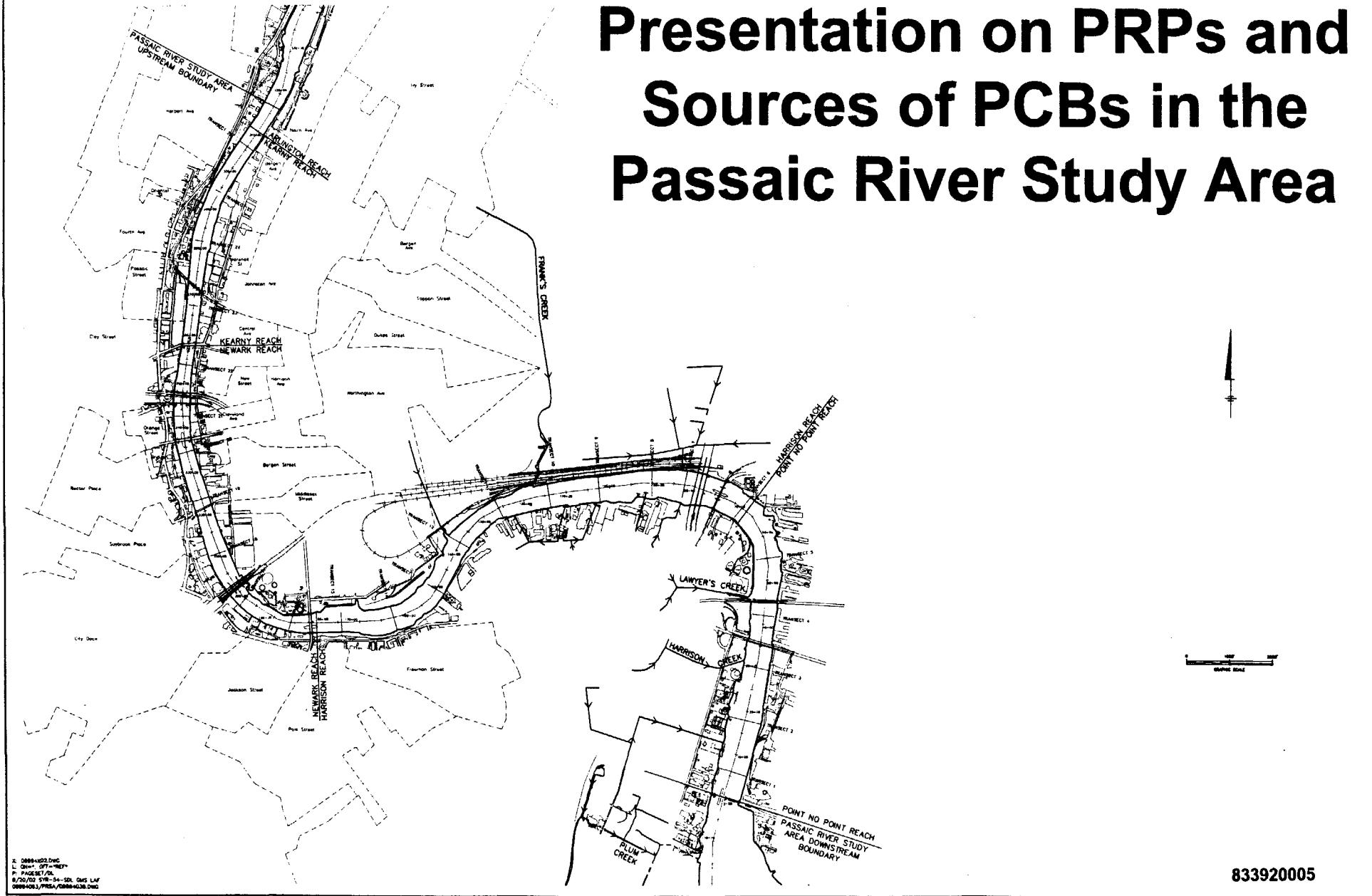




PRPs and Sources of PCBs in the Passaic River Study Area

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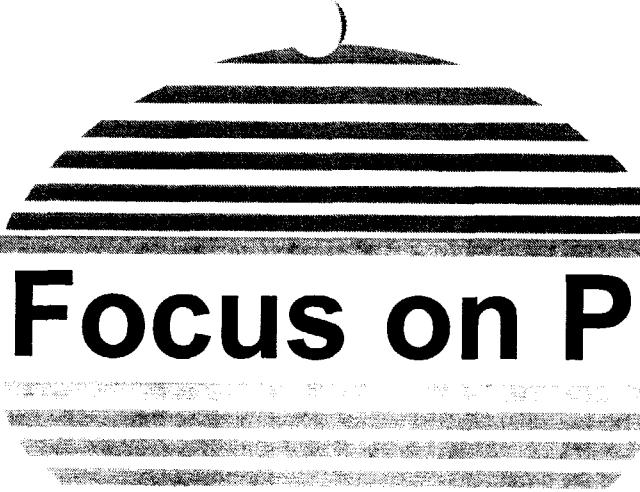
Presentation on PRPs and Sources of PCBs in the Passaic River Study Area





Facts Regarding PCBs in the PRSA

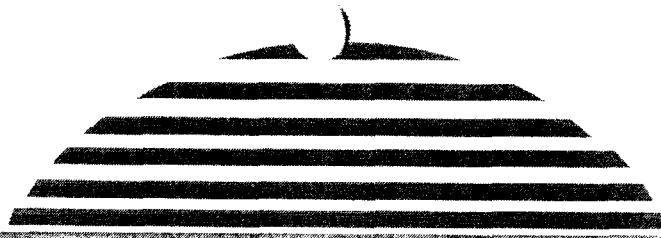
- PRSA sediments contain elevated concentrations of PCBs.
- Numerous potential sources of PCBs to PRSA sediments have been identified – these “PRPs” include historical users and handlers of PCBs and PCB-contaminated products.
- PCB-contaminated soil and/or groundwater exist(s) at many of these PRPs’ upland locations.
- Many of these PRP locations have historical and/or present day discharge pathways to the PRSA.
- Additional investigation will reveal more PRPs – both within the PRSA as well as the PRRI area.



Why Focus on PCBs?

- PCB contamination of sediments important from a Risk Assessment standpoint.
- Fishing Ban in-place since mid-1980s in Newark Bay complex, including the PRSA.
- Many sources are present.

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Sources of PCBs to the Environment

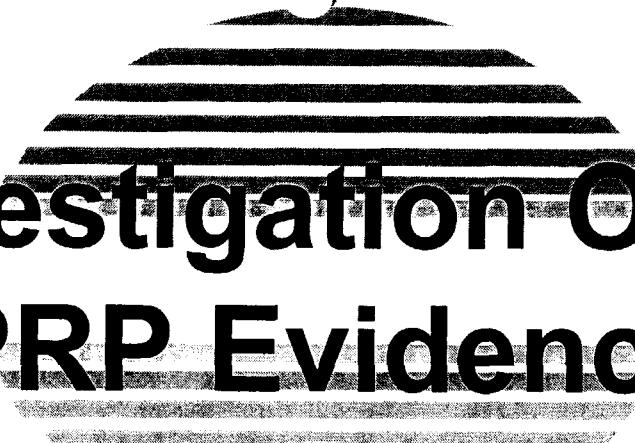
- As manufacturing products, such as Aroclors, for uses including:
 - Electrical capacitors and transformers
 - Vacuum pumps
 - Hydraulic fluids
 - Heat transfer systems
 - Adhesives
 - Paints and inks
 - Plasticizers
 - Cutting oils and de-dusting agents
- As contaminants in recycled oil
- Inadvertent generation, from processes such as:
 - Pigment manufacture
 - Dye manufacture

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PCB Investigation Context

- Focus to date only on PRSA – to assist USEPA in identifying PRPs for PCB contamination.
- Presented to USEPA on 18 December 2001.
- Future focus on the PRRI area will yield additional PRPs.
- Evidence more readily available regarding PCB sources than dioxin sources – most sites are typically sampled for PCBs, but not dioxins.

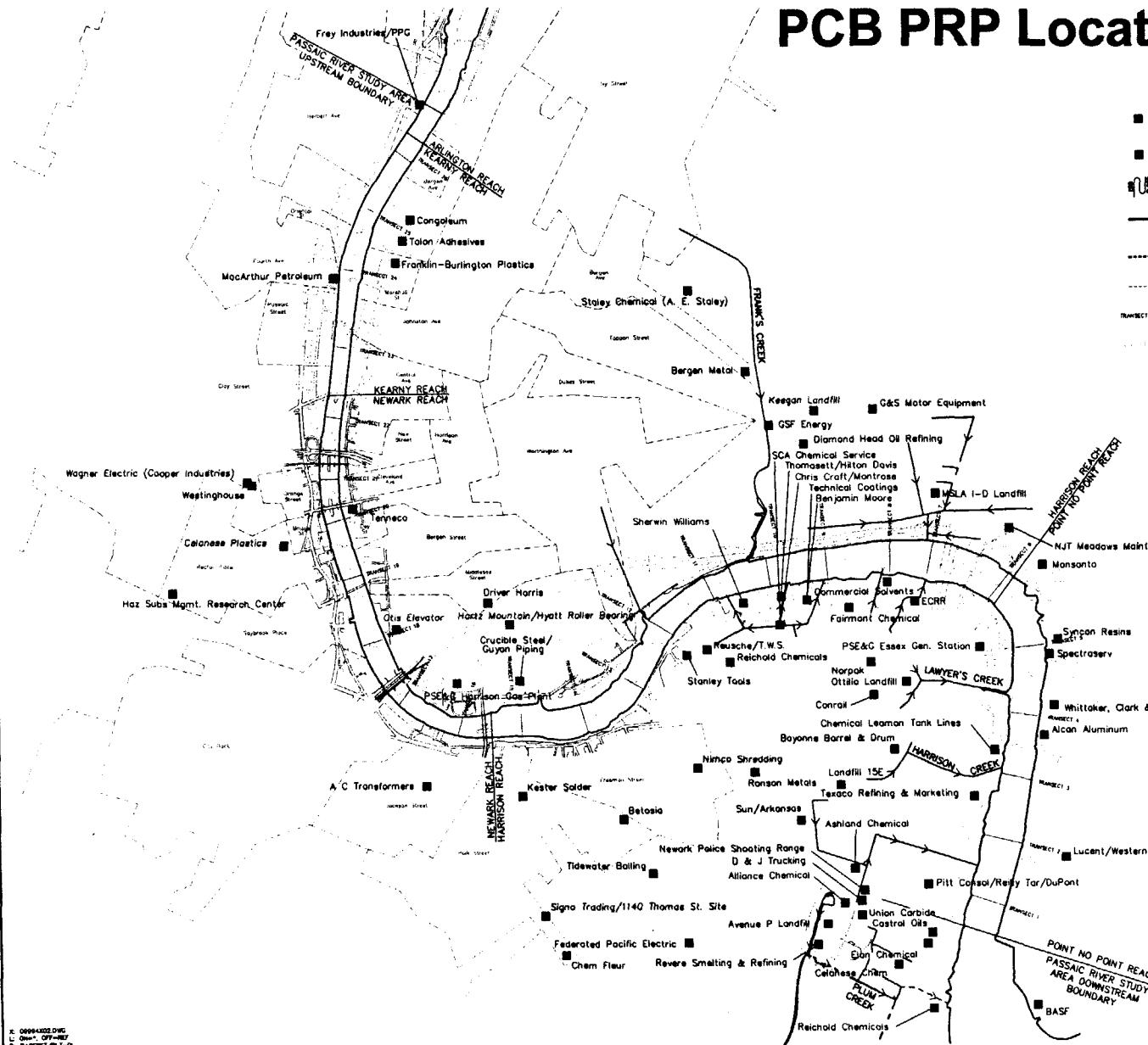


PCB Investigation Overview: PRP Evidence

- Gathered evidence on approximately 75 PRPs.
- PRPs identified are: users/handlers of PCBs, operators of sites with PCB contamination, or entities using processes known to inadvertently generate PCBs.
- Identified from publicly available records, including:
 - USEPA enforcement and compliance records
 - NJDEP site remediation records and files
 - local city records
 - product sales records
 - PRP responses to CERCLA 104(e) requests.

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PCB PRP Locations in the PRSA



LEGEND:

- PRSA PRPs NOTICED BY EPA AS OF SEPTEMBER 2002
- PRP LOCATIONS
- TIDALLY INFLUENCED FLOW
- TRIBUTARY OR DISCHARGE ROUTE TO THE PASSAIC RIVER
- UNDERGROUND FLOW ROUTE
- - - CSO DISTRICT
- 1995 SEDIMENT SAMPLING TRANSECT
- USACE RIVER STATIONING

PRP:
A.C. Transformers
Alcan Aluminum
Alliance Chemical
Ashland Chemical
Avenue P Landfill
BASF
Bayonne Barrel & Drum
Benzene Source
Bergen Metal
Betosia
Celanese Chem
Celanese Plastic
Chem Fleur
Chemical Leamor Tank Lines
Chris Craft/Monrose
Commercial Solvents
Congoleum
Cookster Steel/Guyon Piping
D & J Trucking
Diamond Head Oil Refining
Driver Morris
ECRR
Elon Chemical
Fairmont Chemical
Federated Pacific Electric
Franklin-Burlington Plastics
Frey Industries/PPG
GAS Motor Equipment
GSE Energy
Hartz Mountain/Hyatt Roller Bearing
Haz Subs Mgmt. Research Center
Keegan Landfill
Kester Solder
Landfill 15E
Lucent/Western Electric (AT&T)
MacArthur Petroleum
Monsanto
MSLA 1-D Landfill
Newark Police Shooting Range
Nimco Shredding
NJT Meadows Maintenance
Norpak
Ottie Landfill
P&G Essex Gen. Station
Pitt Conso/Rely Tar/DuPont
PSC&G Gas Plant
PSC&G Newark Gas Plant
Reichold Chemicals
Rausche/T.W.S.
Revere Smelting & Refining
Ronson Metals
SCA Chemical Service
Sherwin Williams
Sigco Trading/1140 Thomas St. Site
Spectraserv
St. Louis Chemical (A. E. Staley)
Stanley Tools
Sun/Arkansas
Syncon Resins
Talon Adhesives
Technical Coatings
Tenneco
Texaco Refining & Marketing
Tidewater Balling
Union Carbide Castrol Oils
Whittaker, Clark & Daniels
Wagner Electric (Cooper Industries)
Westinghouse

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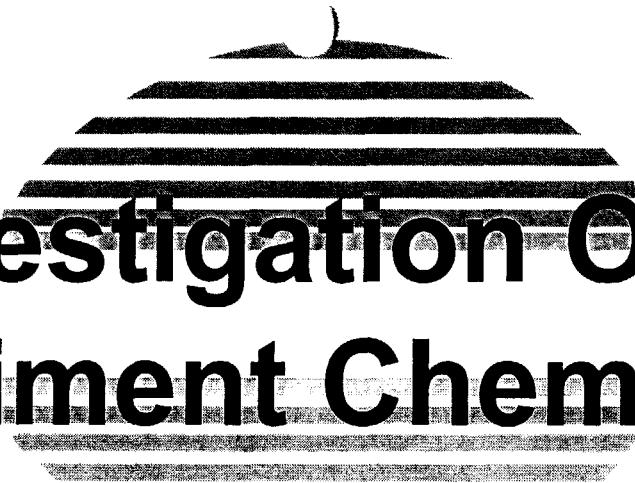
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PCB Investigation Overview: PRP Evidence

- Evaluated evidence for each PRP location to identify historical or present day discharge pathways to the PRSA.
- Compared evidence for each PRP location to the PRSA sediment chemistry near the site's discharge pathway(s).

PCB Investigation Overview: Sediment Chemistry

- PRSA sediment chemistry was reviewed to identify areas of peak concentrations of PCBs.
- Both Aroclors and dioxin-like congeners were utilized:
 - Aroclors historically utilized in sampling of upland sources.
 - Dioxin-like congeners are utilized to assess risk in sediments.



PCB Investigation Overview: Sediment Chemistry

- PCB Aroclors considered are:

1221

1248

1260

1242

1254

- Dioxin-like PCB congeners considered are:

BZ77

BZ118

BZ157

BZ189

BZ105

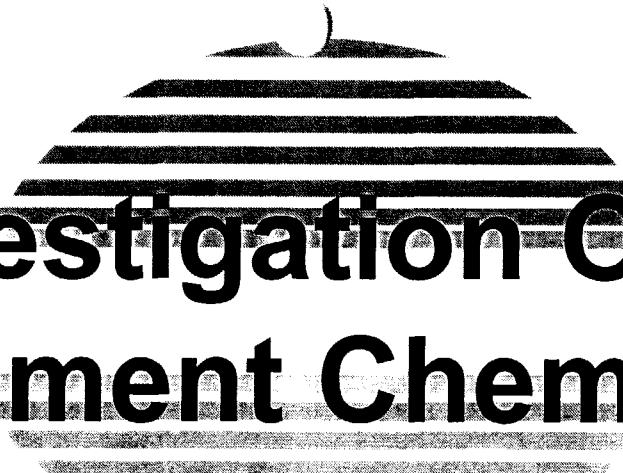
BZ126

BZ167

BZ114

BZ156

BZ169

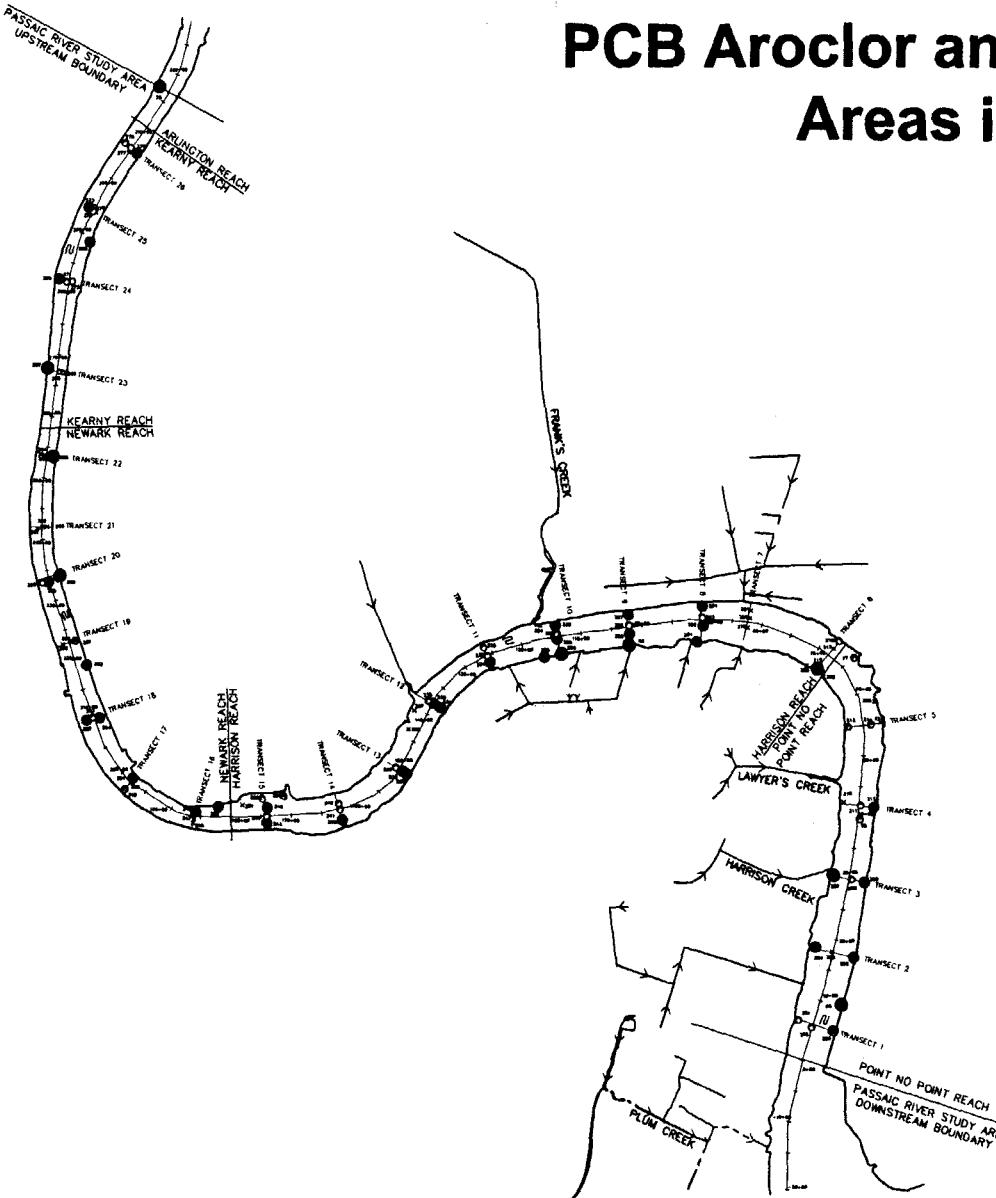


PCB Investigation Overview: Sediment Chemistry

- Grouped sediment data for each Aroclor and congener by:
 - Highest individual measurement
 - Top 5% of detected concentrations
 - Top 25% of detected concentrations
- Each sampling location is a “core” – typically representing 3 to 6 individual sampling depth ranges.
- The sample locations were evaluated as to their proximity to PCB sources, and the sample depth ranges were evaluated to help approximate the period of PCB discharge.

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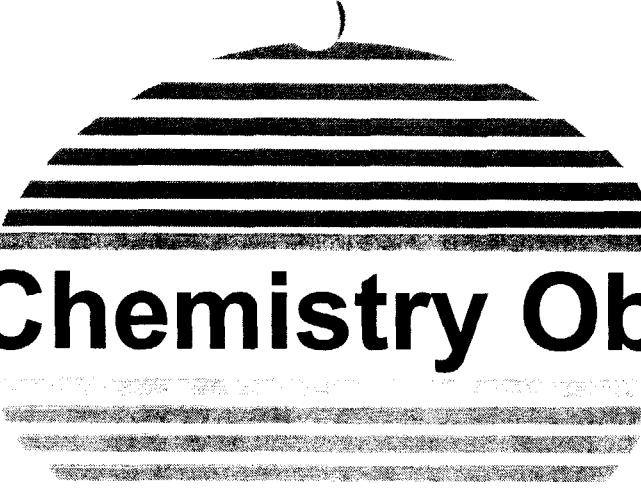
PCB Aroclor and Congener Source Areas in the PRSA



LEGEND:

- TIDALLY INFLUENCED FLOW
- HIGHEST PCB AROCLOR OR CONGENER CONCENTRATION
- TOP 5% PCB AROCLOR OR CONGENER CONCENTRATIONS
- TOP 25% PCB AROCLOR OR CONGENER CONCENTRATIONS. INCLUDES ALL SIGNIFICANT CONCENTRATIONS FROM PCB AROCLORS AND CONGENERS WITH LOW OCCURRENCES OF HITS.
- RI CORE BORING LOCATION
- TRIBUTARY OR DISCHARGE ROUTE TO THE PASSAIC RIVER
- UNDERGROUND FLOW ROUTE
- TRANSECT • 1995 SEDIMENT SAMPLING TRANSECT
- USACE RIVER STATIONING

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Sediment Chemistry Observations

- There is ubiquitous PCB contamination of PRSA sediments.
- Despite the wide-spread nature of this contamination, PRPs can be identified.
- Further investigation will yield additional PRPs.

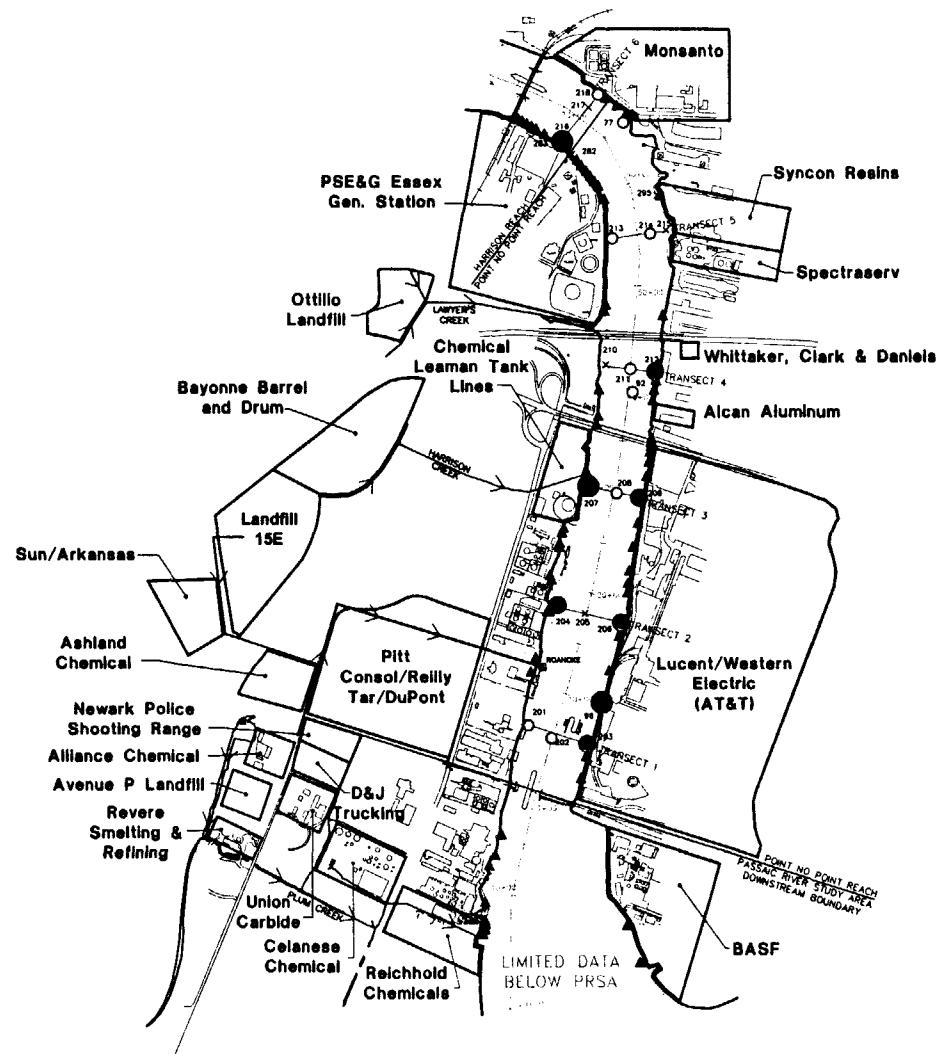


Reach-by-Reach Presentation

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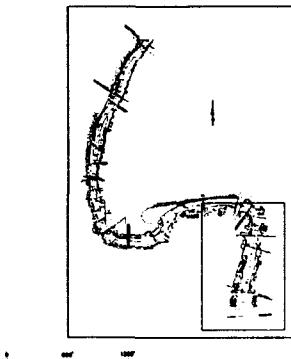
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PCB PRP Locations and PCB Source Areas in Point No Point Reach of the PRSA

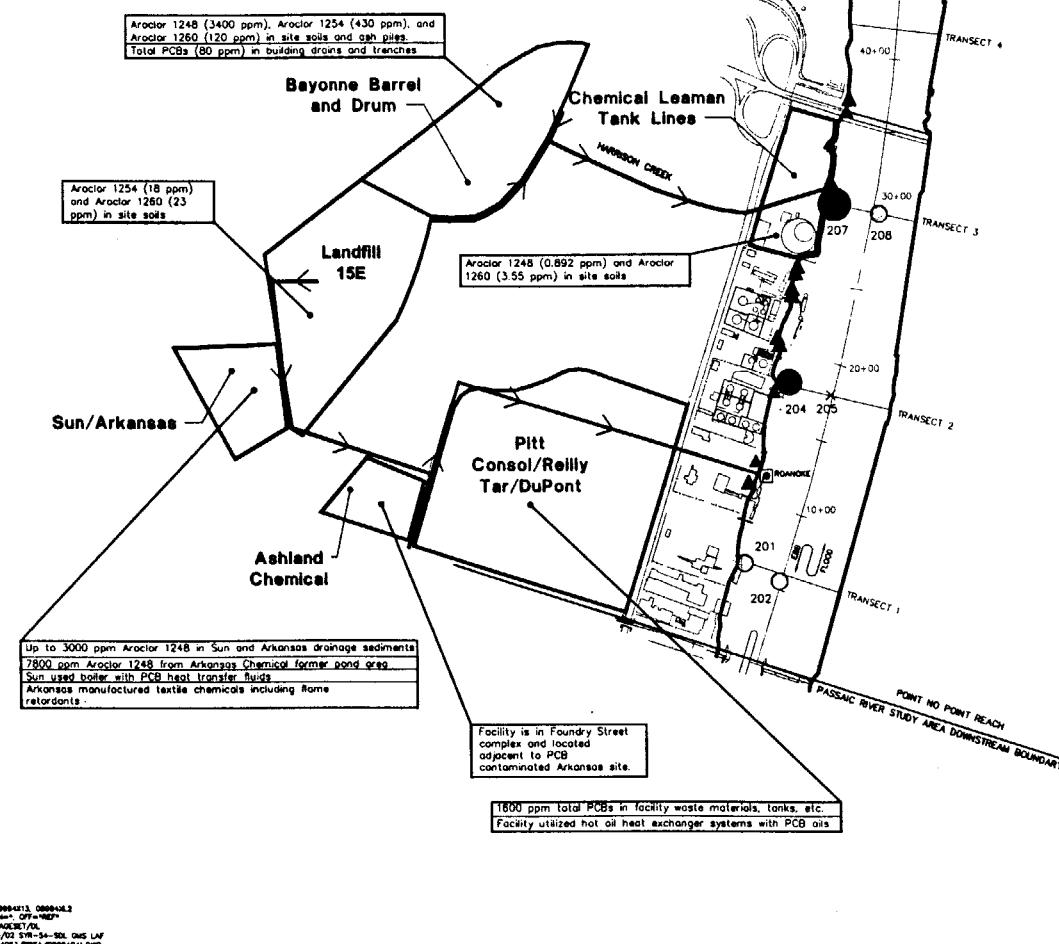


LEGEND:

- TIDALLY INFLUENCED FLOW
- STORM SEWER OUTFALL
- CSO OUTFALL
- HISTORIC AND PRESENT DAY OUTFALLS
- PRP FACILITY OUTLINES
- HIGHEST PCB AROCLOR OR CONGENER CONCENTRATION
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- RI CORE BORING LOCATION
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- 1995 RI SEDIMENT SAMPLING TRANSECT
- USACE RIVER STATIONING



PCB PRP Locations and PCB Source Areas Along Point No Point Reach Western Riverbank



Core 201

Sample	Aroclor	Conc. (ppb)	Congener	Conc. (ppb)
20101A	1248	3880	B2105	0.078
20106A	1248	2040	B2118	230
			B2105	1.5

Core 202

Sample	Aroclor	Conc. (ppb)	Congener	Conc. (ppb)
20205E	1248	80	B2105	170
			B2157	5
20206E	1248	30	B2105	67
			B2118	220
			B2158	22
			B2159	3.5

Core 204

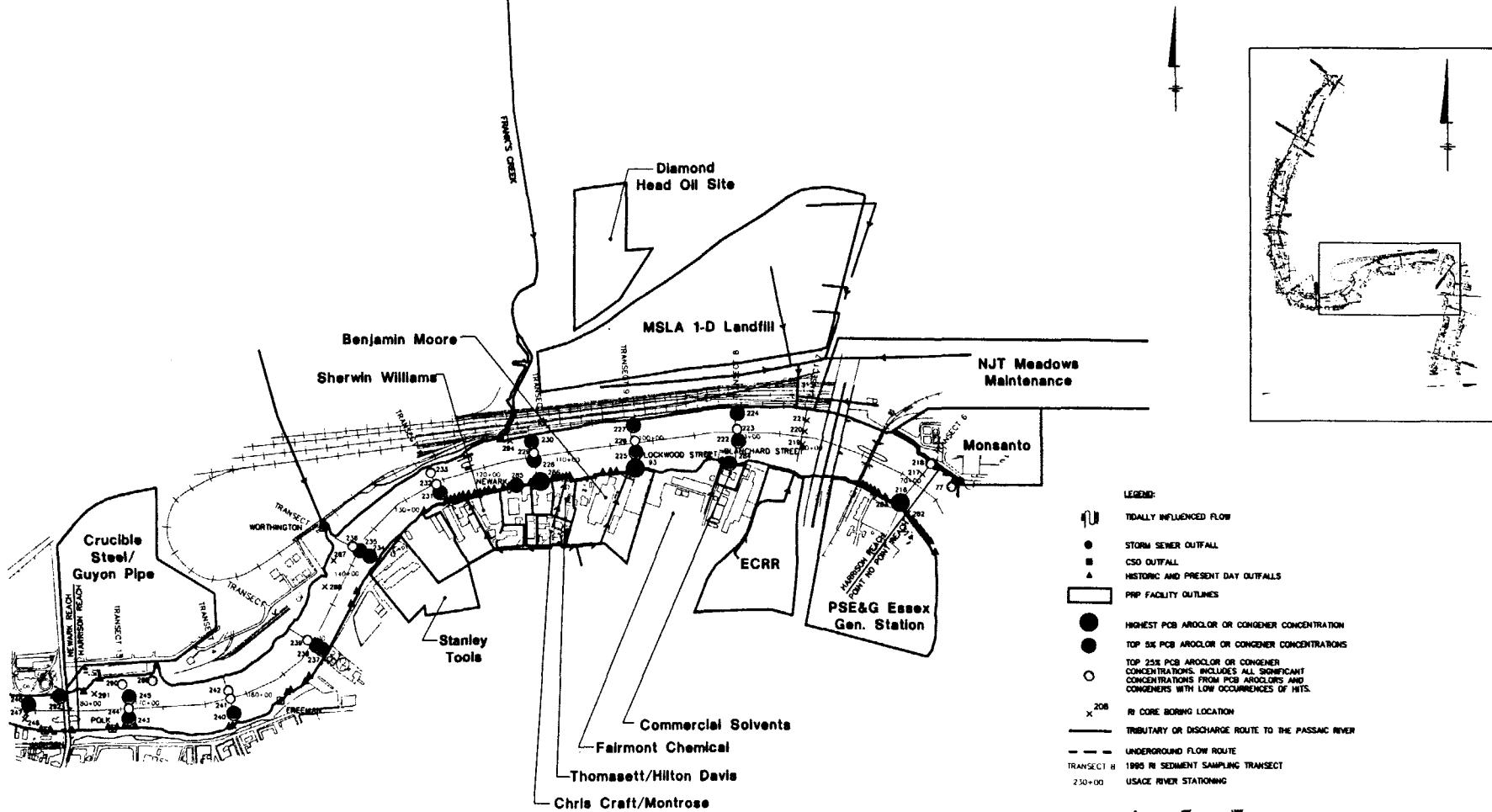
Sample	Aroclor	Conc. (ppb)	Congener	Conc. (ppb)
20405A	1248	4910	B2177	68
	1254	2040	B2105	100
			B2114	9.9
			B2118	205
			B2154	74
			B2157	4.7
			B2167	30
			B2189	1.7
20408A	1248	1550	B2177	55
			B2105	550
			B2114	58
			B2118	10100
			B2154	125
			B2157	15000
			B2167	10000
			B2189	1000

Core 206

Sample	Aroclor	Conc. (ppb)	Congener	Conc. (ppb)
20602A	1248	2570	B2177	62
			B2105	98
			B2114	8.8
			B2118	250
			B2157	5.7
			B2169	0.063

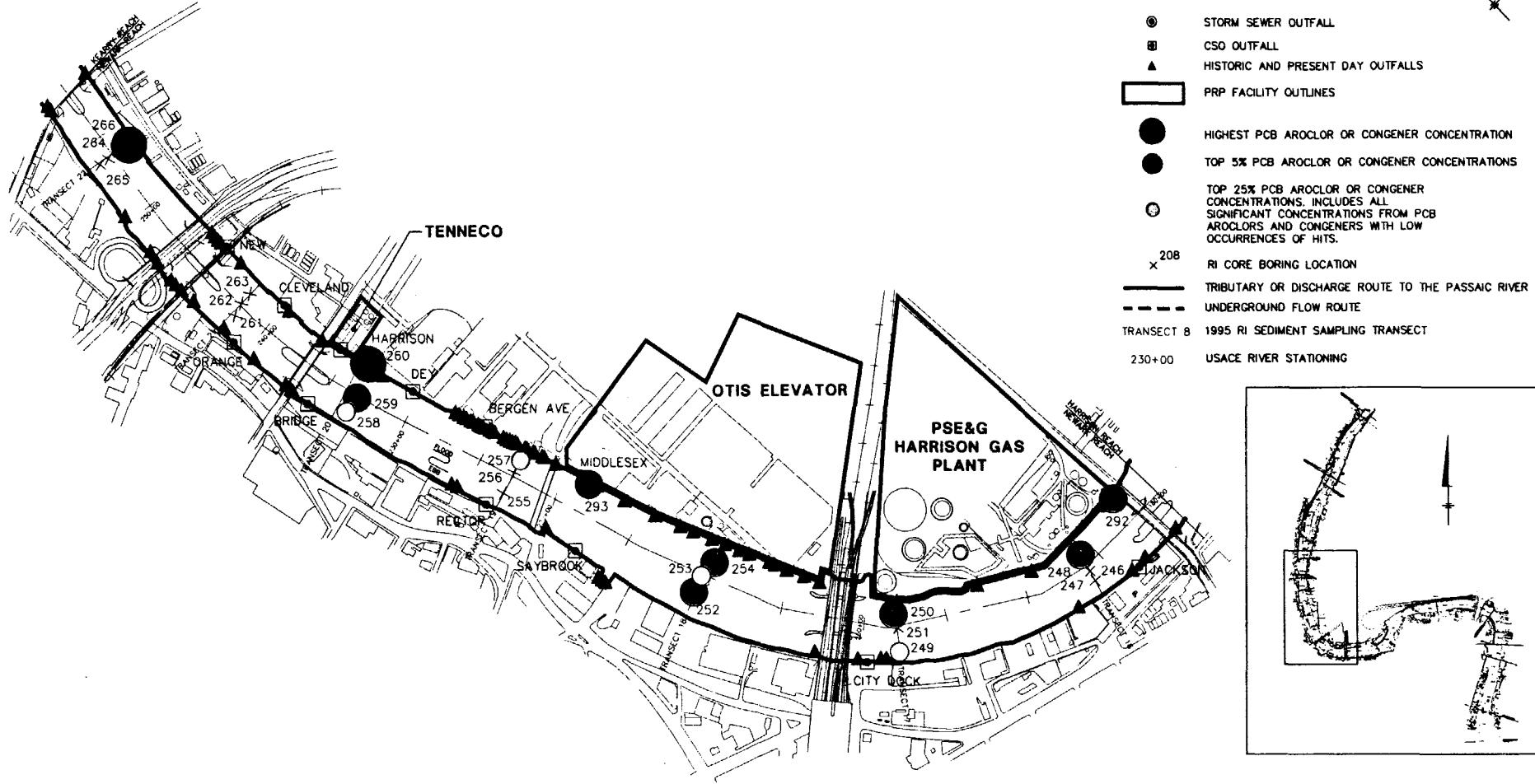
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PCB PRP Locations and PCB Source Areas in Harrison Reach of the PRSA



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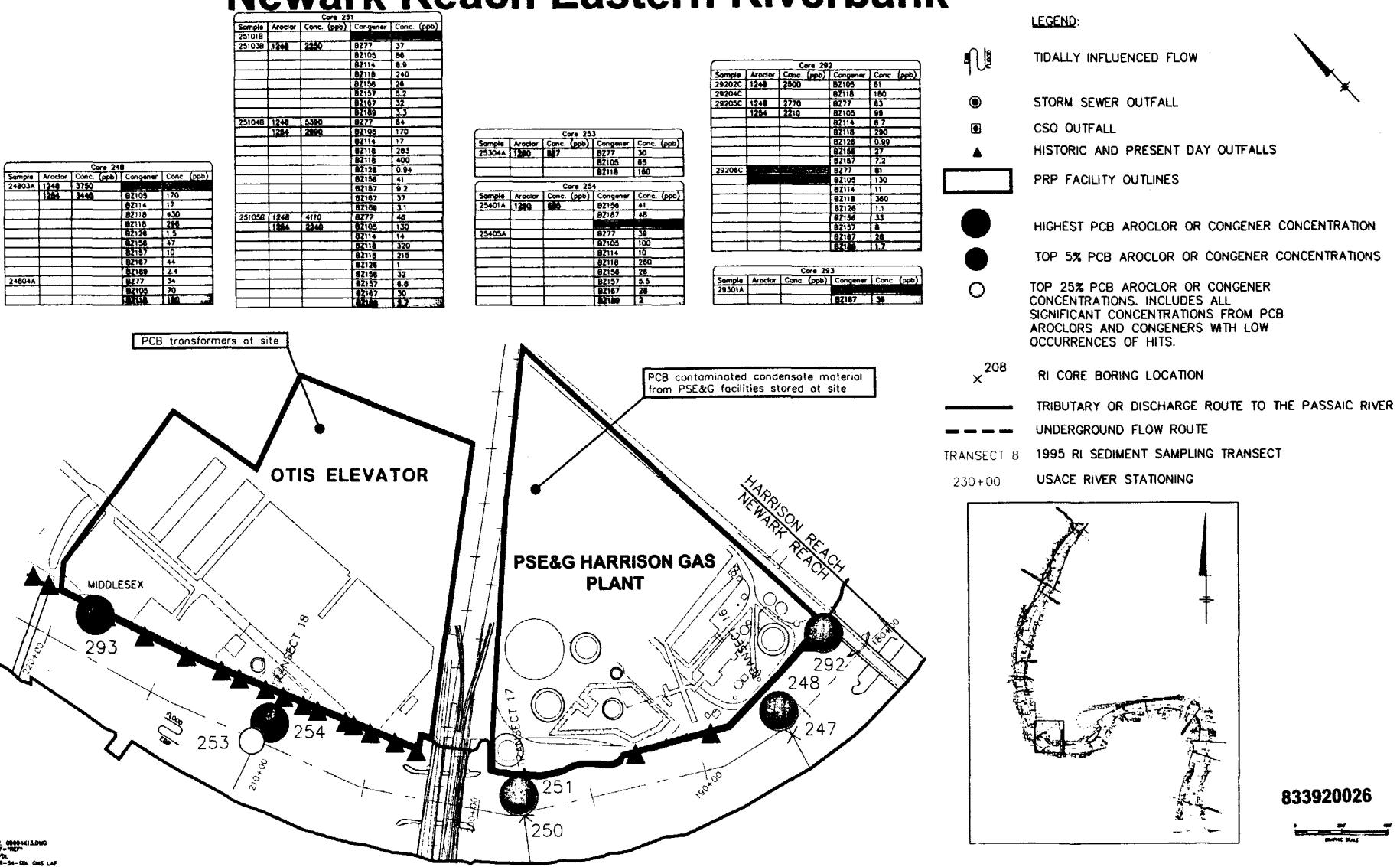
PCB PRP Locations and PCB Source Areas in Newark Reach of the PRSA



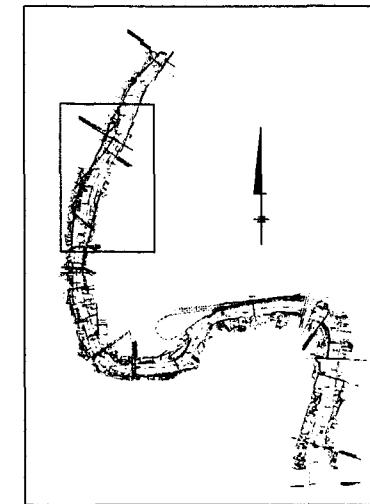
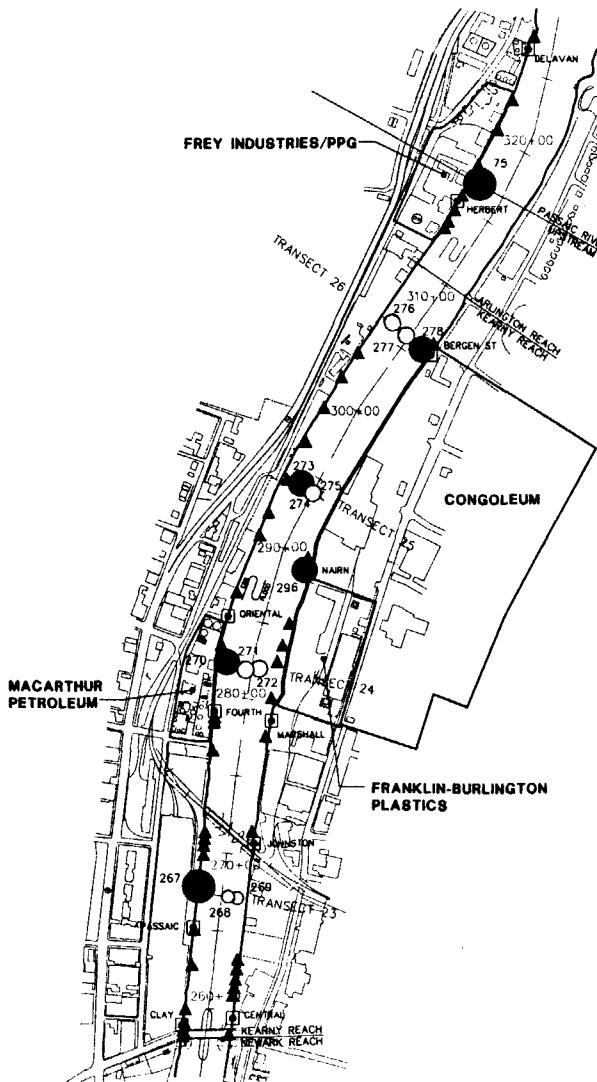
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QMB-OF-0027
P. PAGESET/BL
8/20/02 SYR-5A-SOL QMB LAF
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PCB PRP Locations and PCB Source Areas Along Newark Reach Eastern Riverbank



PCB PRP Locations and PCB Source Areas in Kearny and Arlington Reaches of the PRSA



LEGEND:

- TIDALLY INFLUENCED FLOW
- STORM SEWER OUTFALL
- CSO OUTFALL
- HISTORIC AND PRESENT DAY OUTFALLS
- PRP FACILITY OUTLINES
- HIGHEST PCB AROCLOR OR CONGENER CONCENTRATION
- TOP 5% PCB AROCLOR OR CONGENER CONCENTRATIONS
- TOP 25% PCB AROCLOR OR CONGENER CONCENTRATIONS. INCLUDES ALL SIGNIFICANT CONCENTRATIONS FROM PCB AROCLORS AND CONGENERS WITH LOW OCCURRENCES OF HITS.
- RI CORE BORING LOCATION
- TRIBUTARY OR DISCHARGE ROUTE TO THE PASSAIC RIVER
- UNDERGROUND FLOW ROUTE
- TRANSECT 8 1995 RI SEDIMENT SAMPLING TRANSECT
- 230+00 USACE RIVER STATIONING

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1 100' BRIDGE SPAN

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L 08994302 08994313
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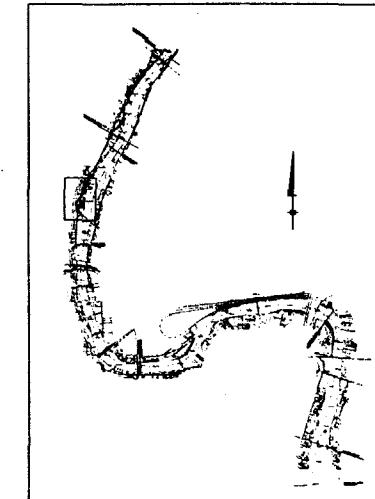
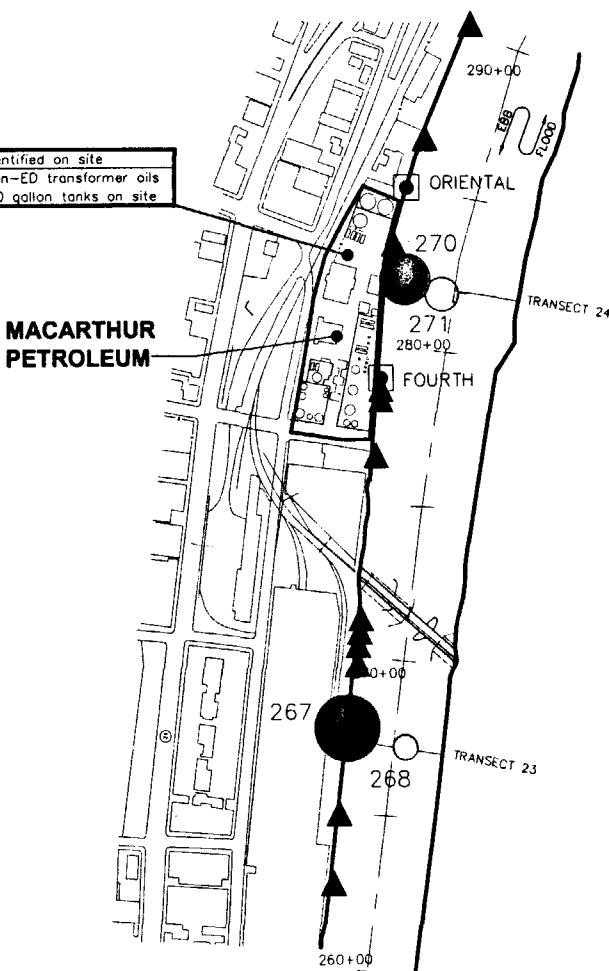
PCB PRP Locations and PCB Source Areas Along Kearny Reach Western Riverbank

Core 267				
Sample	Aroclor	Conc. (ppb)	Congener	Conc. (ppb)
26701A	B277	63		
	B2105	32		
	B2114	10		
	B2118	240		
	B2169	1.7		
26702A	1264	3430	B277	29
	B2118	284		

Core 268				
Sample	Aroclor	Conc. (ppb)	Congener	Conc. (ppb)
26802A	1262	6150	B2105	94
1264	3430	93	B2114	9.8
	B2118	250		
	B2118	167		
	B2156	32		
	B2157	6.2		
	B2167	42		
	B2169	2.9		
27003A	B277	63		
	B2105	33		
	B2114	9.3		
	B2118	247		
	B2128	250		
	B2156	29		
	B2167	34		
27004A	B277	321		
	B2105	1.6		
	B2156	44		
	B2157	8.7		
	B2169	2.2		
27006A	B277	120		
	B2105	120		
	B2114	120		
	B2118	120		
	B2128	14		
	B2156	1.8		
	B2169	2.6		

Core 270				
Sample	Aroclor	Conc. (ppb)	Congener	Conc. (ppb)
27001A	1262	6150	B2105	94
1264	3430	93	B2114	9.8
	B2118	250		
	B2118	167		
	B2156	32		
	B2157	6.2		
	B2167	42		
	B2169	2.9		
27003A	B277	63		
	B2105	33		
	B2114	9.3		
	B2118	247		
	B2128	250		
	B2156	29		
	B2167	34		
27004A	B277	321		
	B2105	1.6		
	B2156	44		
	B2157	8.7		
	B2169	2.2		
27006A	B277	120		
	B2105	120		
	B2114	120		
	B2118	120		
	B2128	14		
	B2156	1.8		
	B2169	2.6		

Core 271				
Sample	Aroclor	Conc. (ppb)	Congener	Conc. (ppb)
27102A	1260	1020	B277	34
27103A	1264	2080	B2105	87
	B2114	7.2		
	B2118	190		
	B2167	30		
	B2169	1.8		
27104A	1248	4860	B277	59
1264	3430	82105	120	
	B2114	120		
	B2118	330		
	B2128	289		
	B2156	1.3		
	B2157	31		
	B2167	6.2		
	B2169	44		
	B2180	1.8		

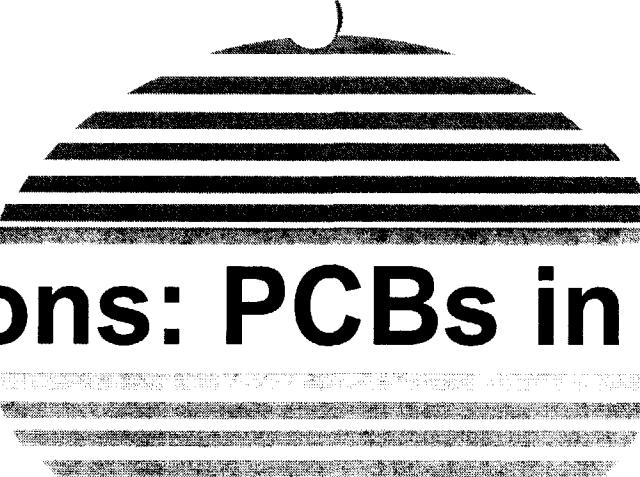


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LEGEND:

- TIDALLY INFLUENCED FLOW
- STORM SEWER OUTFALL
- CSO OUTFALL
- HISTORIC AND PRESENT DAY OUTFALLS
- PRP FACILITY OUTLINES
- HIGHEST PCB AROCLOR OR CONGENDER CONCENTRATION
- TOP 5% PCB AROCLOR OR CONGENDER CONCENTRATIONS
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- X CORE BORING LOCATION
- TRIBUTARY OR DISCHARGE ROUTE TO THE PASSAIC RIVER
- - - UNDERGROUND FLOW ROUTE
- TRANSECT 8 1985 SEDIMENT SAMPLING TRANSECT



Conclusions: PCBs in the PRSA

- PRSA sediments contain elevated concentrations of PCBs.
- Numerous potential sources of PCBs to PRSA sediments have been identified – these “PRPs” include historical users and handlers of PCBs and PCB-contaminated products.
- PCB-contaminated soil and/or groundwater exist(s) at many of these PRPs’ upland locations.
- Many of these PRP locations have historical and/or present day discharge pathways to the PRSA.
- Additional investigation will reveal more PRPs – both within the PRSA as well as the PRRI area.

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PCB PRP Locations in the PRSA

LEGEND:

■ PRSA PRPs NOTICED BY EPA AS OF SEPTEMBER 2002

■ PRP LOCATIONS

■ TIDALLY INFLUENCED FLOW

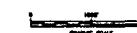
— TRIBUTARY OR DISCHARGE ROUTE TO THE PASSAIC RIVER

----- UNDERGROUND FLOW ROUTE

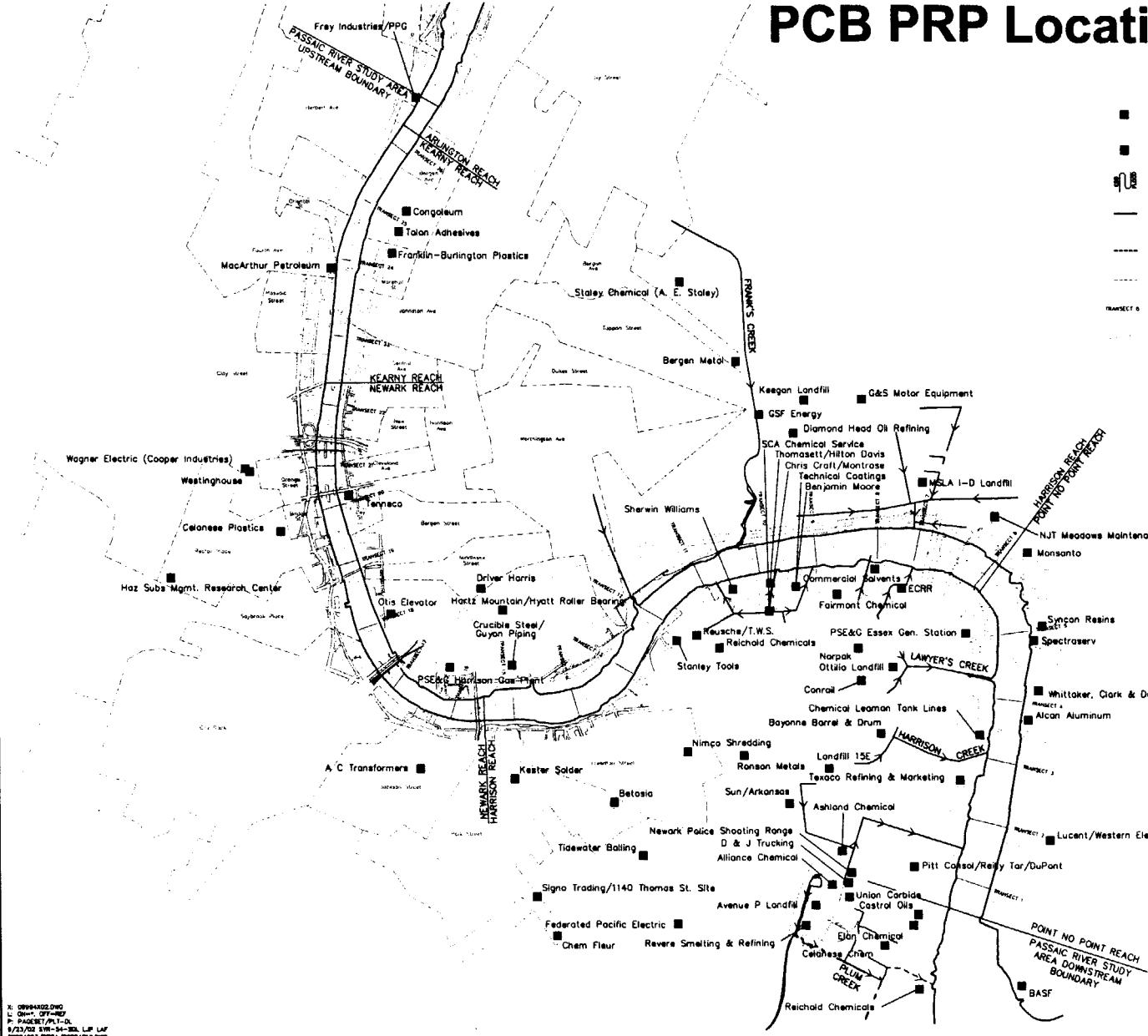
— CSO DISTRICT

TRANSECT ■ 1995 SEDIMENT SAMPLING TRANSECT

USACE RIVER STATIONING



PRP:
A.C. Transformers
Alcan Aluminum
Alliance Chemical
Ashland Chemical
Avenue P Landfill
BASF
Bayonne Barrel & Drum
Benjamin Moore
Bergen Metal
Bethel
Celanese Chem
Celanese Plastics
Chem Fleur
Chemical Leamor Tank Lines
Chris Craft/Montrose
Commercial Solvents
Congoleum
Conval
Diamond Steel/Guyon Piping
D & J Trucking
Diamond Head Oil Refining
Driver Harris
ECRR
Ebasco Chemical
Fairmont Chemical
Federated Pacific Electric
Franklin-Burlington Plastics
Frigidaire Industries/PPG
GAS Equipment
GSF Energy
Hartz Mountain/Hyatt Roller Bearing
Haz Subs Mgmt. Research Center
Kearny Landfill
Kester Solder
Landfill 15E
Lucas/Western District (AT&T)
MacArthur Petroleum
MSPA I-D Landfill
Newark Police Shooting Range
Nimco Shredding
NJIT Meadows Maintenance
Norpak
Ottie Elevator
Ortilio Landfill
Pitt Consol/Rally Tar/DuPont
PSEG Essex Gen. Station
PSEG Harrison Gas Plant
Reichold Chemicals
Reusche/T.W.S.
Revere Smelting & Refining
Ronson Metals
SAC Chemical Services
Sherwin Williams
Sigro Trading/1140 Thomas St. Site
Sigro Trading
Stedt Chemical (A. E. Staley)
Stanley Tools
Sun/Arkansas
Syncon Resins
Talon Adhesives
Technical Coatings
Tenneco
Tidewater Baiting
Union Carbide
Union Carbide Control Oils
Union Carbide Refining & Marketing
Thomasset/Anthon Doyle
Tidewater Baiting
Whitaker, Clark & Daniels

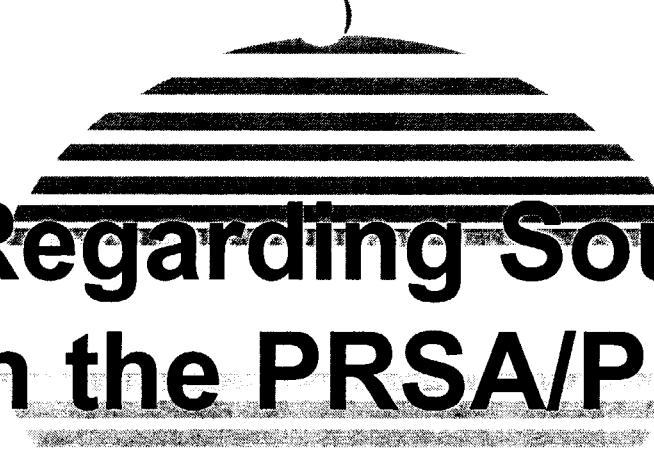




For Next Time:

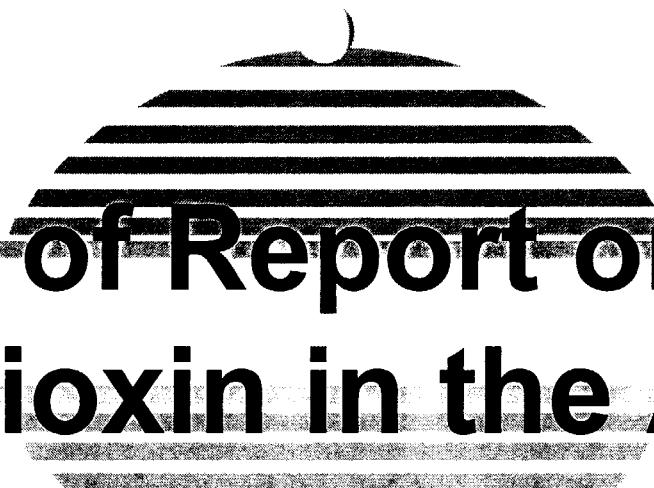
**Presentation on Sources of Dioxin
in the PRRI Area**

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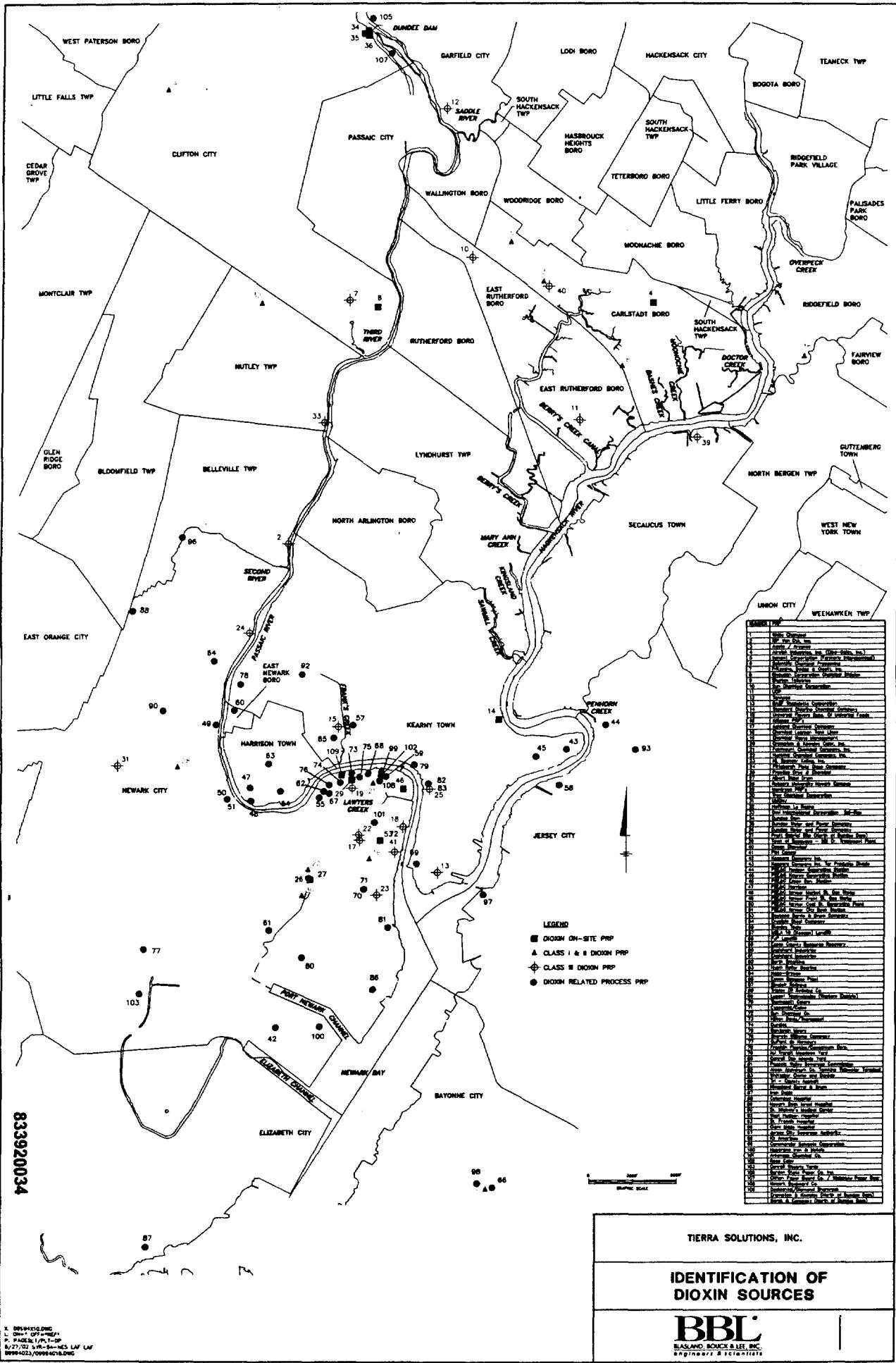
Facts Regarding Sources of Dioxin in the PRSA/PRRI Area

- There exist PRPs that handled products and employed processes utilizing chlorinated phenols – just like the former Diamond site.
- There exist PRPs that handled chemicals and employed processes identified by USEPA as associated with the formation of dioxins.
- Sampling for dioxins at these PRP locations has been limited; but dioxins were detected where sampling was conducted .
- Many of these PRP locations have historical and/or present day discharge pathways to the PRRI area.
- Additional investigation is required.



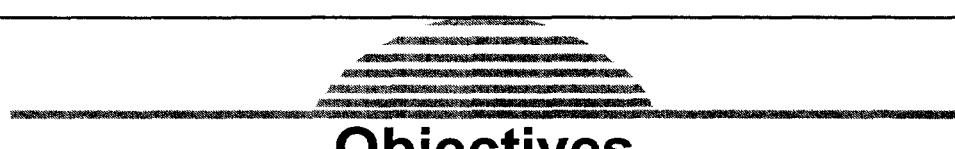
Overview of Report on Sources of Dioxin in the Area

- Provides background on the formation of dioxins.
- Identifies more than 100 PRPs in the area associated with actual or likely dioxin generation.
- Provides evidence regarding 5 of these PRPs.





Habitat Characterization



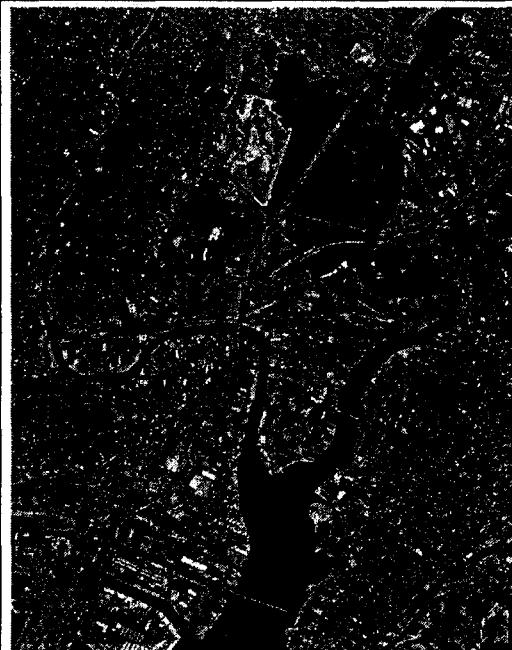
Objectives

- Identify key habitats that remain in the PRSA
- Characterize/quantify shoreline habitats in the PRSA
- Delineate intertidal mudflats

Methods

- Visual/videotape survey of shorelines throughout PRSA in Fall 1999 and Spring 2000
 - Low and high tide surveys
 - Classify/quantify shorelines into four categories — aquatic vegetation, bulkhead, riprap, mixed vegetation

3



4

Lower Passaic River Landscape



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Shoreline Habitat Classification Categories

Category	Description
Aquatic vegetation	Represents shoreline habitats composed of emergent wetland plant species such as <i>Spartina alterniflora</i> or <i>Phragmites</i> . Areas of aquatic vegetation often occur as narrow bands of vegetation near the top of the intertidal zone, typically with intertidal mudflat below.
Bulkhead	Consists of horizontal or vertical wood timbers, metal sheet pile, or large stone blocks constructed to form a vertical face perpendicular to the water surface
Riprap	Includes cobble to boulder-sized stone and/or concrete rubble placed along the shoreline on a sloped bank
Mixed vegetation	Represents areas with aquatic vegetation interspersed (laterally and/or longitudinally) with riprap and/or bulkhead. Areas of riprap shoreline with significant over-hanging riparian vegetation were also included as mixed vegetation to acknowledge the minor contribution to aquatic habitat provided by the adjacent riparian vegetation.

5



Typical Bulkhead

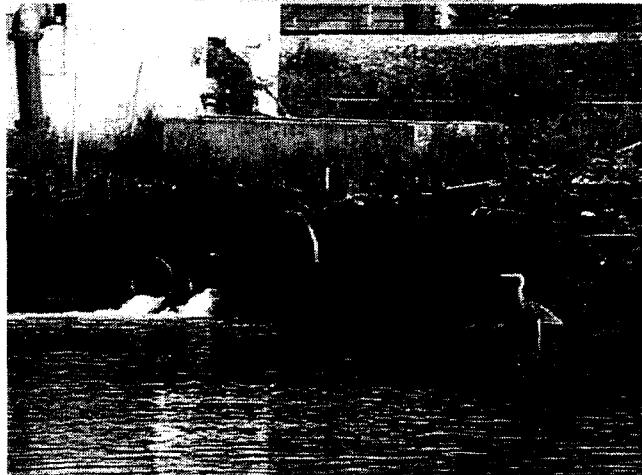


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Bulkhead, Riprap, Outfalls



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Mixed Vegetation/Mudflat



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Shoreline Habitat Characterization for the PRSA — Point-No-Point Reach

Shoreline Habitat Type	Right Bank ^a		Left Bank ^b	
	Linear Feet	Percent of Total	Linear Feet	Percent of Total
Bulkhead	1,219	16%	4,994	63%
Riprap	4,128	54%	2,873	37%
Mixed vegetation ^c	883	12%	0	0%
Aquatic vegetation	1,407	18%	0	0%
Total shoreline (feet)	7,637		7,867	

Notes:

- ^a Right bank facing downstream (e.g., western/southern shoreline).
- ^b Left bank facing downstream (e.g., eastern/northern shoreline).
- ^c Mixed vegetation refers to areas of aquatic/upland vegetation interspersed with riprap or bulkhead and areas of riprap shoreline with significant overhanging riparian vegetation.



Shoreline Habitat Characterization for the PRSA — Harrison Reach

Shoreline Habitat Type	Right Bank ^a		Left Bank ^b	
	Linear Feet	Percent of Total	Linear Feet	Percent of Total
Bulkhead	4,524	39%	3,131	25%
Riprap	4,508	38%	4,037	32%
Mixed vegetation ^c	2,171	19%	3,409	27%
Aquatic vegetation	519	4%	1,917	15%
Total shoreline (feet)	11,722		12,494	

Notes:

- ^a Right bank facing downstream (e.g., western/southern shoreline).
- ^b Left bank facing downstream (e.g., eastern/northern shoreline).
- ^c Mixed vegetation refers to areas of aquatic/upland vegetation interspersed with riprap or bulkhead and areas of riprap shoreline with significant overhanging riparian vegetation.



Shoreline Habitat Characterization for the PRSA – Newark Reach

Shoreline Habitat Type	Right Bank ^a		Left Bank ^b	
	Linear Feet	Percent of Total	Linear Feet	Percent of Total
Bulkhead	6,860	81%	5,973	77%
Riprap	1,562	19%	1,796	23%
Mixed vegetation ^c	0	0%	0	0%
Aquatic vegetation	0	0%	0	0%
Total shoreline (feet)	8,422		7,769	

Notes:

^a Right bank facing downstream (e.g., western/southern shoreline).

^b Left bank facing downstream (e.g., eastern/northern shoreline).

^c Mixed vegetation refers to areas of aquatic/upland vegetation interspersed with riprap or

^c bulkhead and areas of riprap shoreline with significant overhanging riparian vegetation.

Shoreline Habitat Characterization for the PRSA – Kearny Reach

Shoreline Habitat Type	Right Bank ^a		Left Bank ^b	
	Linear Feet	Percent of Total	Linear Feet	Percent of Total
Bulkhead	4,802	90%	3,214	62%
Riprap	526	10%	800	15%
Mixed vegetation ^c	0	0%	1,189	23%
Aquatic vegetation	0	0%	0	0%
Total shoreline (feet)	5,328		5,203	

Notes:

^a Right bank facing downstream (e.g., western/southern shoreline).

^b Left bank facing downstream (e.g., eastern/northern shoreline).

^c Mixed vegetation refers to areas of aquatic/upland vegetation interspersed with riprap or

^c bulkhead and areas of riprap shoreline with significant overhanging riparian vegetation.

Shoreline Habitat Characterization for the PRSA — Arlington Reach

Shoreline Habitat Type	Right Bank ^a		Left Bank ^b	
	Linear Feet	Percent of Total	Linear Feet	Percent of Total
Bulkhead	573	89%	0	0%
Riprap	70	11%	30	4%
Mixed vegetation ^c	0	0%	655	96%
Aquatic vegetation	0	0%	0	0%
Total shoreline (feet)	643		685	

Notes:

- ^a Right bank facing downstream (e.g., western/southern shoreline).
- ^b Left bank facing downstream (e.g., eastern/northern shoreline).
- ^c Mixed vegetation refers to areas of aquatic/upland vegetation interspersed with riprap or bulkhead and areas of riprap shoreline with significant overhanging riparian vegetation.

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Shoreline Habitat Characterization for the PRSA — Cumulative Total

Shoreline Habitat Type	Right Bank ^a		Left Bank ^b		Total Shoreline	
	Linear Feet	Percent of Total	Linear Feet	Percent of Total	Linear Feet	Percent of Total
Bulkhead	17,978	53%	17,312	51%	35,290	52%
Riprap	10,794	32%	8,536	28%	20,330	30%
Mixed vegetation ^c	3,054	9%	5,253	15%	8,307	12%
Aquatic vegetation	1,926	6%	1,917	6%	3,843	6%
Total shoreline (feet)	33,752		34,018		67,770	

Notes:

- ^a Right bank facing downstream (e.g., western/southern shoreline).
- ^b Left bank facing downstream (e.g., eastern/northern shoreline).
- ^c Mixed vegetation refers to areas of aquatic/upland vegetation interspersed with riprap or bulkhead and areas of riprap shoreline with significant overhanging riparian vegetation.

14



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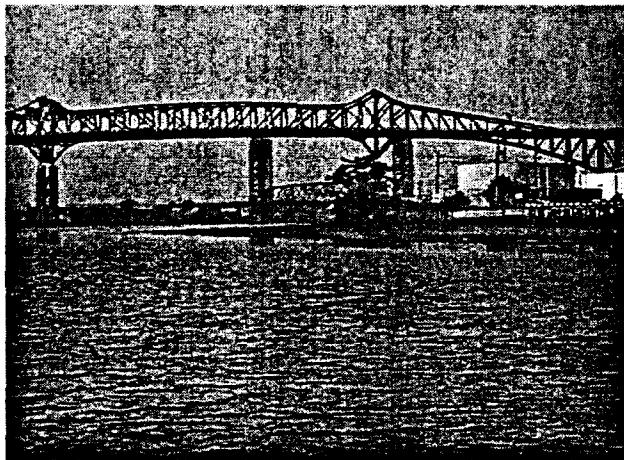
Key Habitats in PRSA

- Intertidal mudflats (although very degraded)
- Frank's Creek confluence area
 - Limited *Spartina alterniflora* stand
- Lawyer's Creek confluence area
 - Mixed *Phragmites australis* and *Spartina* stand

15



Typical Mudflat Area



16



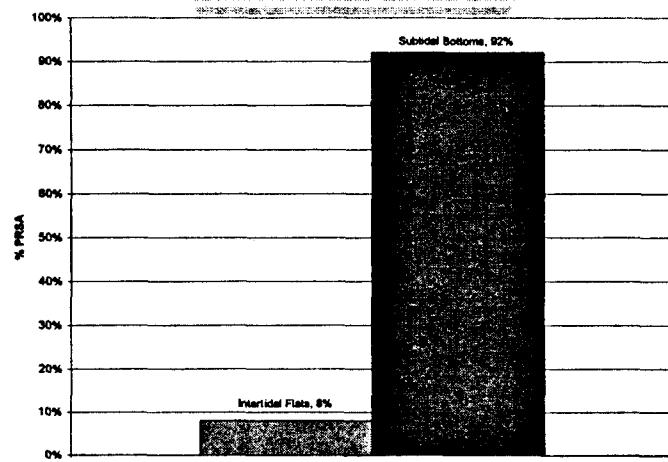
Typical Mudflat Area



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River Bottom Habitat



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Estimates of Historical Wetland Losses in the Newark Bay Estuary

Year	Acres	Cumulative Percent Loss
Pre-1816 ^a	24,466	--
1870	18,166	26
1905	15,790	36
1932	11,968	51
1940	11,180	54
1954	8,738	64
1966	5,574	77
1976	3,570	85
1989	3,058	88
1997	2,921	88

^a Based on sum of mapped wetlands in 1870 and reported wetlands losses for period of 1816 through 1867.

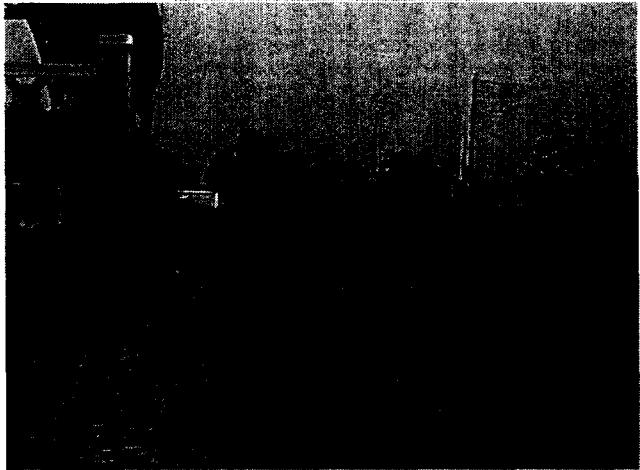


Estimated Losses of Historical Rivers, Creeks, and Tributaries in the Lower Passaic River and Newark Bay

River/Creek	Estimated Length Lost (mi)
Bound Creek and Tributaries	18.1
Maple Island Creek and Tributaries	13.2
First River and Tributaries	6.0
Unnamed Passaic Tributary Creeks	0.7
Kearny Marsh Tributaries	1.2
Great Meadow Brook and Tributaries	6.3
Oyster Creek and Tributaries	2.3
Upper Newark Bay Tributaries	10.9
Other Newark Bay Tributaries	20.2
Total Lost	76.6



Lawyer's Creek Confluence with PRSA



21



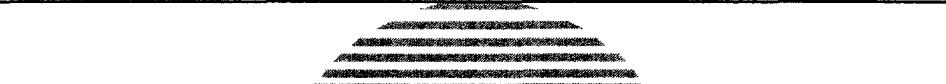
Frank's Creek Confluence with PRSA



22



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Conclusions

- Wetlands limited primarily to degraded intertidal mudflats and fringe vegetation
- Majority of shoreline (>80%) consists of bulkhead and riprap = very little habitat value
- Less than 10% of the shoreline area contains aquatic/wetlands vegetation



Benthic Invertebrate Community Characterization



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Objectives

- Compare structure and composition of benthic invertebrate community in PRSA to Mullica River reference area
- Contrast differences between stations in PRSA
- Conduct sediment quality triad (SQT) assessment



Methods

- 15 PRSA stations/3 reference area stations
- Fall 1999 and Spring 2000 sampling
- 3 replicate samples per station (middle sampling grid)
- Modified Van Veen sampler — biologically active zone (about 0- to 6-inch depth)
- Identification to lowest practicable taxon
- Assessment of community structure/composition metrics

3



Results

- High inter-station variability in both PRSA and reference area
- Seasonal variability
- Many “impacted” stations in PRSA

4



Description of Benthic Invertebrate Community Composition Metrics

Metric	Description
Percent abundance of crustacea	In general, crustacea (particularly amphipods) are largely recognized as taxa that are sensitive to pollutants in aquatic environments. Their presence in samples (expressed as a percent of the total number of species) is considered a good indicator of non-toxic conditions and favorable habitat in the substrate.
Percent pollution-tolerant organisms	Represented mainly by opportunistic oligochaetes that will typically dominate (or co-dominate) samples in stressed aquatic environments. The sum of individuals in these taxa are calculated and expressed as a relative contribution (%) to the total number of individuals in the sample.

7



Benthic Invertebrate Community Structure Metrics

Station	Number of Individuals (Ind/m ²)	Shannon-Wiener Diversity Index					
		Number of Taxa	Pielou's Evenness	Braillouin Diversity	Swartz's Dominance Index	Virginia Province	Biotic Index
<i>PRSA</i>							
1	2,855	4	0.40	0.29	0.39	1	-1.6
2	1,072	4	0.64	0.46	0.63	1	-0.84
3	1,261	7	1.0	0.51	0.98	2	-0.72
4	1,145	3	0.23	0.21	0.22	1	-1.1
5	1,507	4	0.50	0.36	0.50	1	-0.62
6	725	6	0.76	0.43	0.75	1	0.86
7	681	5	0.60	0.38	0.59	1	-0.24
8	754	5	0.79	0.49	0.78	1	-0.042
9	1,290	4	0.71	0.51	0.71	1	-6.8
10	1,087	2	0.069	0.10	0.067	1	-7.0
11	11,913	3	0.71	0.64	0.71	2	-66
12	9,971	2	0.29	0.42	0.29	1	-56
13	217	6	1.6	0.90	1.6	4	0.090
14	1,493	2	0.36	0.52	0.36	1	-9.6
15	1,623	4	0.71	0.51	0.70	2	-9.3
<i>Reference Area</i>							
21	841	8	1.5	0.73	1.5	3	1.7
22	1,609	8	0.73	0.35	0.72	1	1.7
23	101	3	0.95	0.87	0.91	2	-0.73

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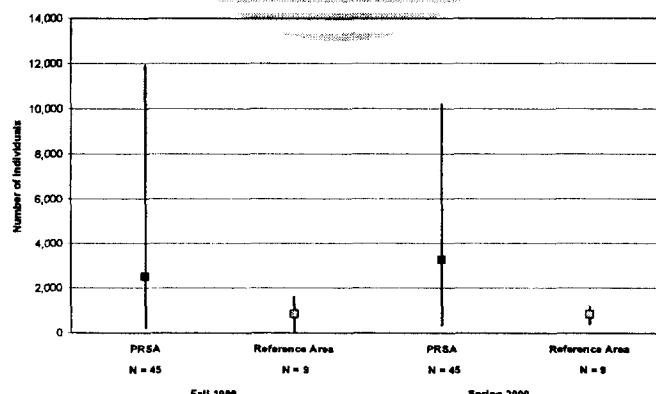
Benthic Invertebrate Community Composition Metrics

Station	Percent Abundance of Crustaceans	Percent Pollution- Tolerant Organisms
	PRSA	
1	8%	2%
2	20%	0%
3	8%	14%
4	4%	0%
5	3%	0%
6	16%	0%
7	4%	2%
8	17%	6%
9	0%	93%
10	0%	99%
11	0%	100%
12	0%	100%
13	27%	13%
14	0%	100%
15	0%	99%
Reference Area		
21	72%	0%
22	94%	0%
23	57%	0%

9



Benthic Invertebrate Community Assessment: Number of Individuals

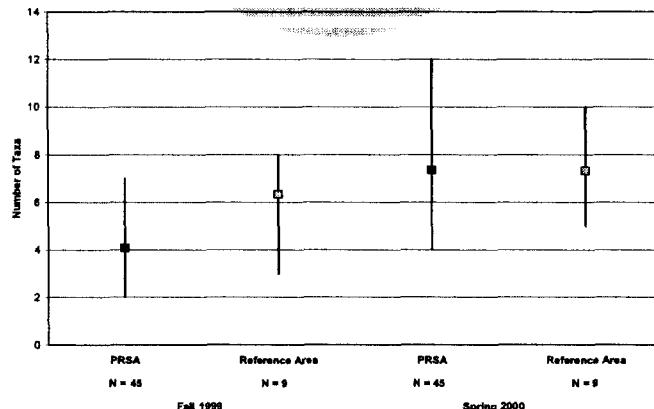


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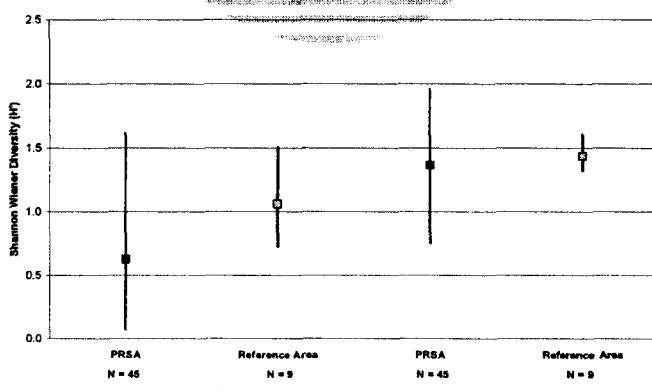
Benthic Invertebrate Community Assessment: Number of Taxa



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Benthic Invertebrate Community Assessment: Shannon-Wiener Diversity (H')

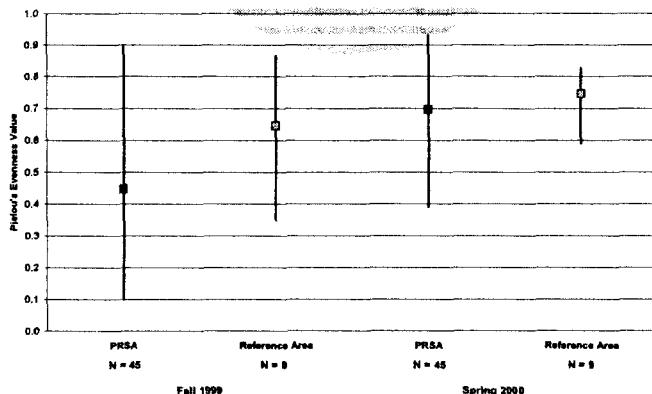


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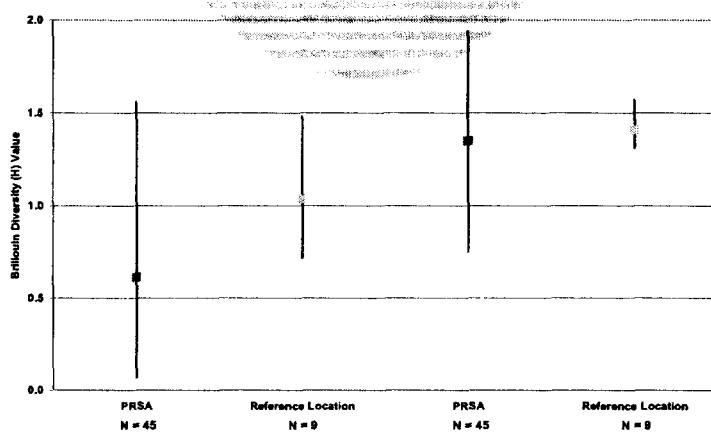
Benthic Invertebrate Community Assessment: Pielou's Evenness



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Brillouin Diversity (H)

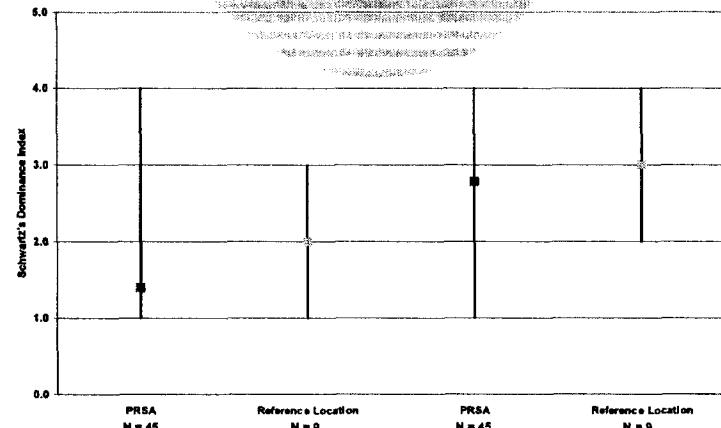


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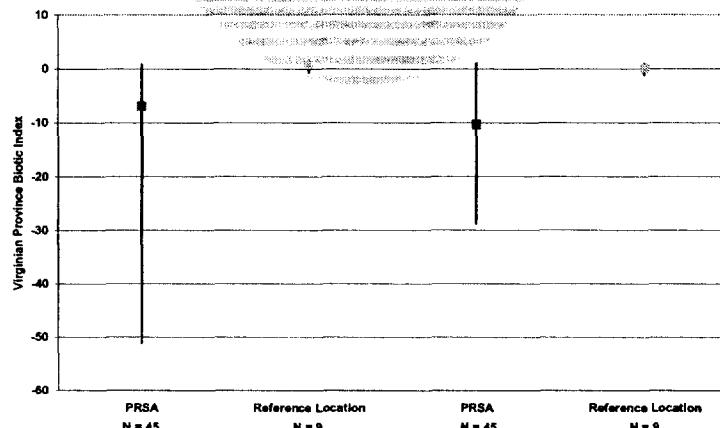
Swartz's Dominance Index



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Virginian Province Biotic Index

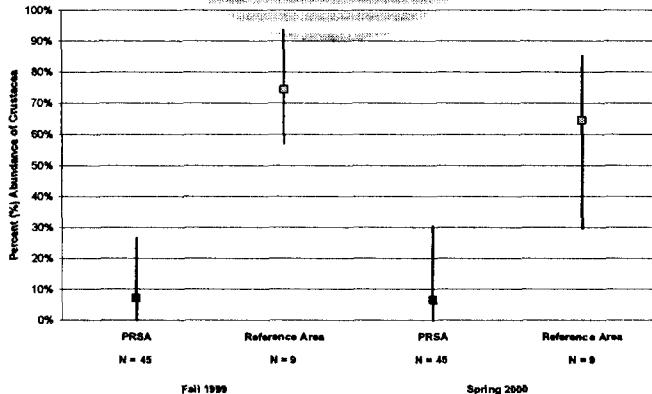


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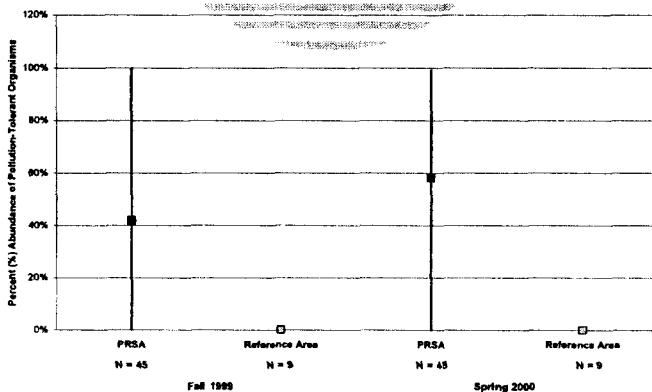
Benthic Invertebrate Community Assessment: Percent Abundance of Crustacea



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Benthic Invertebrate Community Assessment: Percent Abundance of Pollution-Tolerant Organisms



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Classification System for PRSA Benthic Invertebrate Communities

										Virginia Province Index of Biotic Integrity			Abundance of Crustacea		Abundance of Pollution-tolerant Taxa		
		Number of Individuals above Reference		Number of Taxa below Reference		Shannon's H'		Pielou's Evenness below Reference		Brillouin's H below Reference		Swartz Dominance Index below Reference		Reference Area range (<0.73)		Reference Area range (<0.73)	
Poor	Reference Area range (>1,609/m ²) within Reference	Area range (<3)	Area range (<0.73)	Area range (<0.73)	Area range (<0.73)	Area range (<0.72)	Area range (<0.72)	Area range (<1)	Area range (<1)	Area range (<0.73)	Area range (<0.73)	Area range (<1)	Area range (<1)	Area range (<0.73)	Area range (<0.73)	Area range (>0%)	
	Reference Area range (101 - 1,609/m ²) below Reference	within Reference	within Reference	within Reference	within Reference	within Reference	within Reference	within Reference	within Reference	within Reference	within Reference	within Reference	within Reference	within Reference Area range (57 - 94%)	within Reference Area range (0%)	within Reference Area range (0%)	
Good	Reference Area range (<101/m ²) above Reference	Area range (3 - 8)	Area range (0.73 - 1.5)	Area range (0.73 - 0.87)	Area range (0.72 - 0.87)	Area range (0.72 - 1.5)	Area range (0.72 - 1.5)	Area range (1 - 3)	Area range (1 - 3)	Area range (0.73 - 1.7)	Area range (0.73 - 1.7)	Area range (0.73 - 1.7)	Area range (0.73 - 1.7)	Area range (>94%)	Area range (>94%)	Area range (>94%)	
	Excellent	Area range (>8)	Area range (>1.5)	Area range (>0.87)	Area range (>1.5)	Area range (>1.5)	Area range (>1.5)	Area range (>3)	Area range (>3)	Area range (>1.7)	Area range (>1.7)	Area range (>1.7)	Area range (>1.7)	NA	NA	NA	

Note:

- NA - Not applicable

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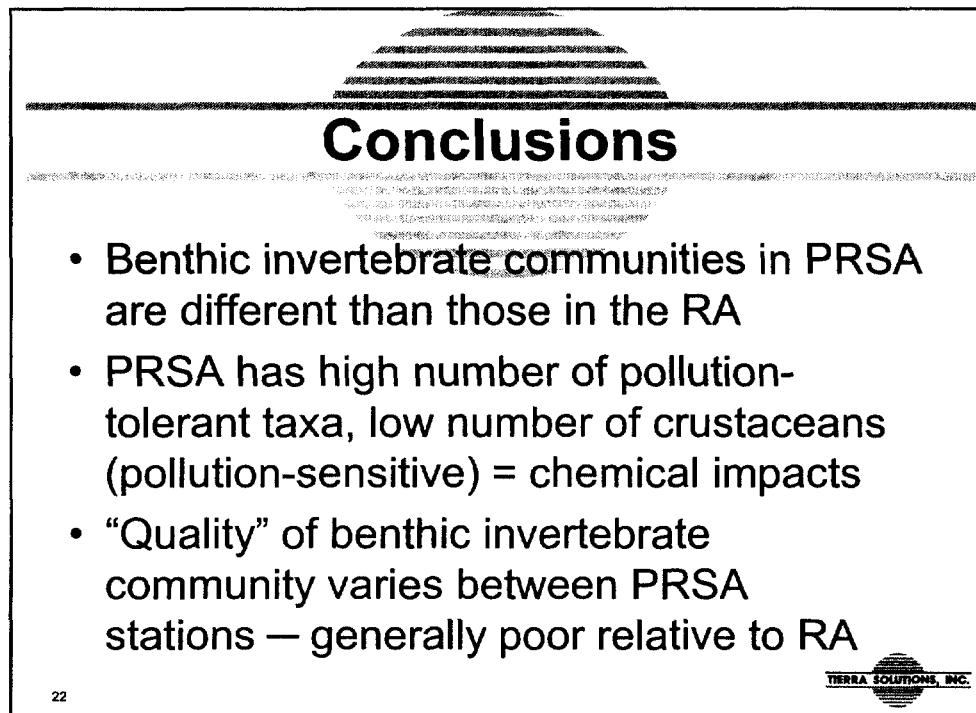
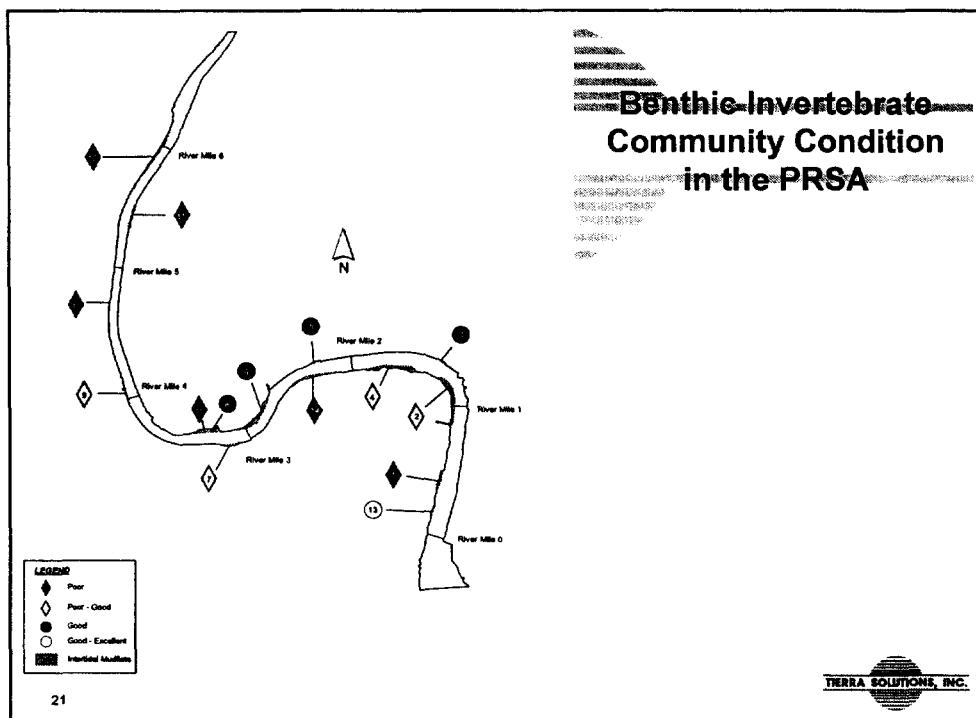
Qualitative Ranks for Each PRSA Station Compared to Reference Area

Metric	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15
No. of Individuals ¹	poor	good													
No. of Taxa	good														
Abundance of Crustaceae	poor														
Abundance of Tolerant Taxa ¹	good														
Pielou's Evenness	good														
Shannon's H'	poor														
Virginia IBI	poor														
Brillouin's H	poor														
Swartz Dominance Index	good														

Note:

¹ For the number of individuals and abundance of tolerant taxa metrics, the following ranks were assigned to each PRSA and Reference Area comparison: a) above reference range = poor; b) within reference range = good; c) below reference range = excellent. For the remaining metrics, the following ranks were assigned for each PRSA/Reference Area comparison: a) above reference area = excellent; b) within reference area = good; c) below reference area = poor

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Fish Community Characterization



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Objectives

- Characterize fish community of PRSA semi-quantitatively on a seasonal basis (Late Summer/Early Fall 1999, Spring 2000)
- Use surveys to confirm/select representative species for contaminant tissue sampling program
- Conduct qualitative pathology investigation on fish not collected for tissue samples



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Methods

- Three target sampling areas/stations in PRSA — lower, middle, and upper river
- Multiple gear types — gill nets, eel traps, minnow traps, crab traps
- Intensive fishing effort in Late Summer/Early Fall 1999 (herein referred to as Fall 1999) and Spring 2000 — driven by tissue targets

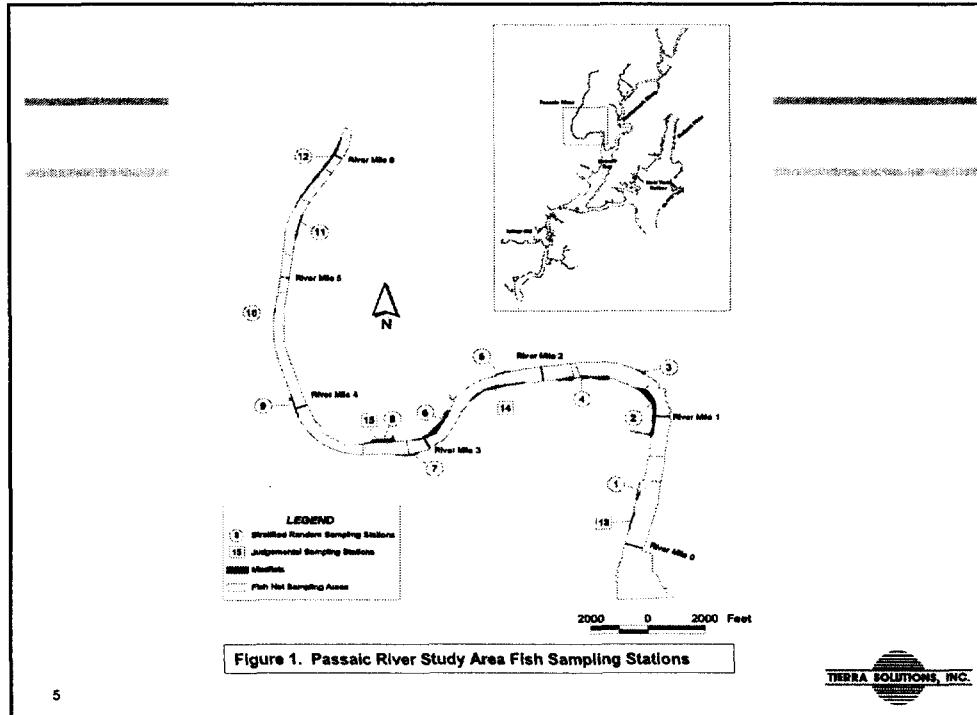
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Methods (cont.)

- Length, weight, and pathology information collected for several species
- Minnow traps set at 15 PRSA stations to collect mummichog tissue samples
- Abundance, dominance, and catch-per-unit-effort (CPUE) calculated
- No reference area for fish community investigation

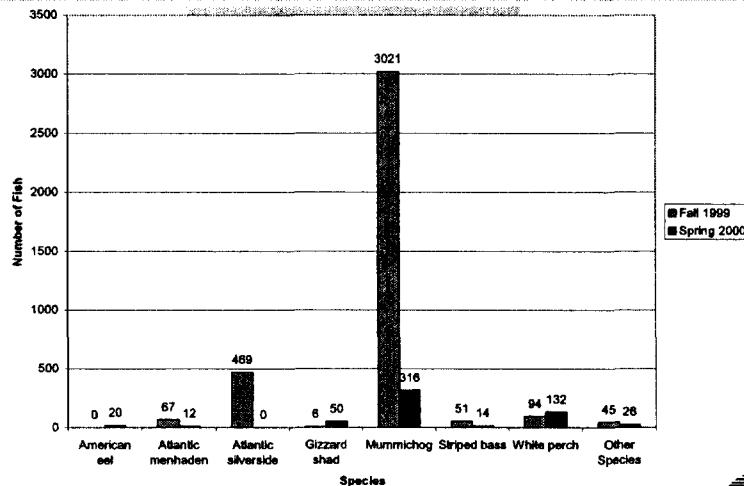




List of Species Caught in PRSA – Fall 1999 and Spring 2000

Common Name	Scientific Name	Fish Caught Fall 1999	Fish Caught Spring 2000
American eel	<i>Anguilla rostrata</i>		X
Atlantic menhaden	<i>Brevoortia tyrannus</i>	X	X
Atlantic silverside	<i>Menidia menidia</i>	X	
Blueback herring	<i>Alosa aestivalis</i>	X	X
Bluefish	<i>Pomatomus saltatrix</i>	X	
Bluegill	<i>Lepomis macrochirus</i>	X	
Brown bullhead	<i>Ameiurus nebulosus</i>		X
Channel catfish	<i>Ictalurus punctatus</i>	X	
Common carp	<i>Cyprinus carpio</i>		X
Gizzard shad	<i>Dorosoma cepedianum</i>	X	X
Green sunfish	<i>Lepomis cyanellus</i>	X	
Inland silverside	<i>Menidia beryllina</i>	X	
Largemouth bass	<i>Micropterus salmoides</i>	X	
Mummichog	<i>Fundulus heteroclitus</i>	X	X
Redear sunfish	<i>Lepomis microlophus</i>	X	
Spotted hake	<i>Urophycis regia</i>		X
Striped bass	<i>Morone saxatilis</i>	X	X
Striped killifish	<i>Fundulus majalis</i>	X	
Summer flounder	<i>Paralichthys dentatus</i>	X	
Weakfish	<i>Cynoscion regalis</i>	X	
White catfish	<i>Ameiurus catus</i>		X
White perch	<i>Morone americana</i>	X	X
White sucker	<i>Catostomus commersoni</i>		X

Number of Fish Caught in the PRSA

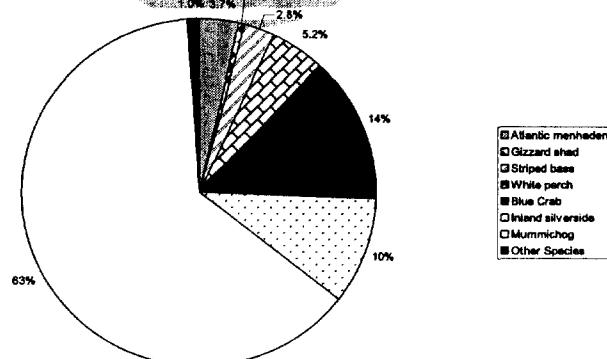


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Percent CPUE Dominance of Fish Caught in the PRSA – Fall 1999

"Other Species" category includes blueback herring, bluefish, bluegill, brown bullhead, channel catfish, common carp, green sunfish, largemouth bass, redear sunfish, spotted hake, striped killifish, summer flounder, weakfish, white catfish, and white sucker.



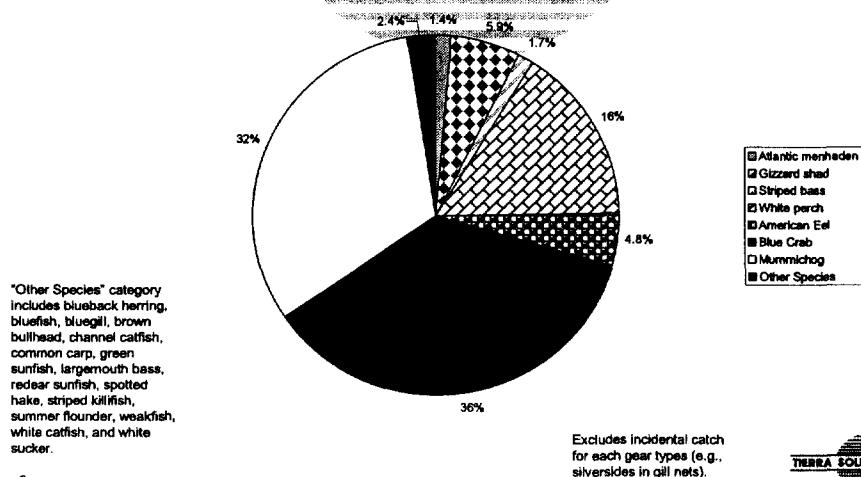
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Excludes incidental catch for each gear types (e.g. silversides in gill nets).

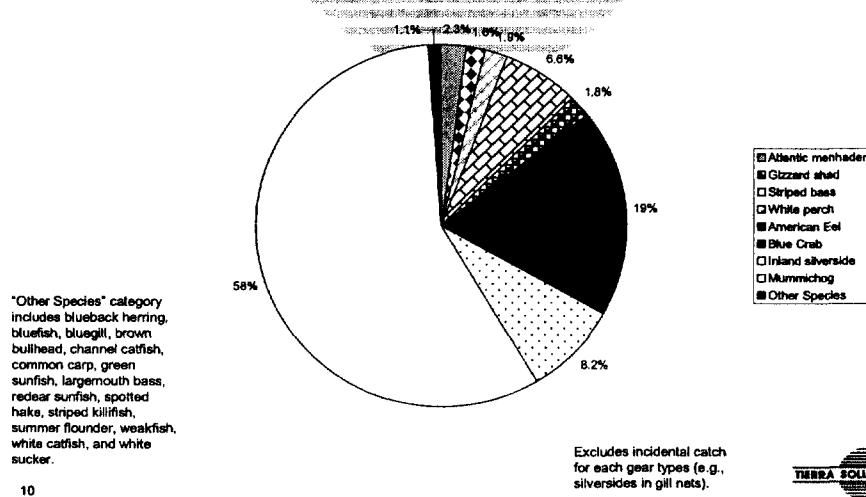
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Percent CPUE Dominance of Fish Caught in the PRSA — Spring 2000

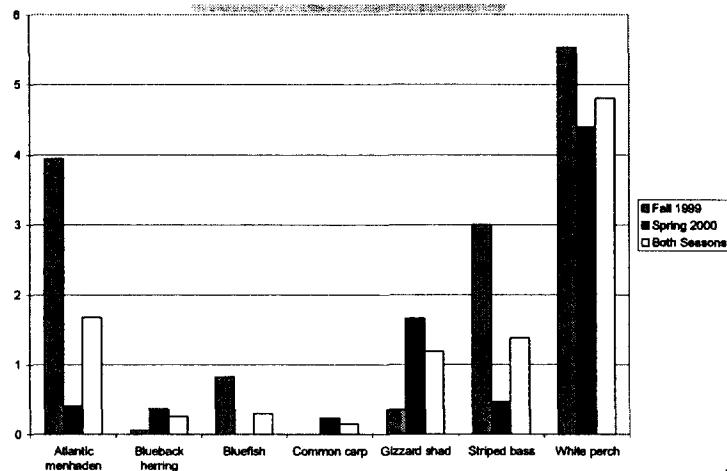


Percent CPUE Dominance of Fish Caught in the PRSA — Combined Fall 1999 and Spring 2000



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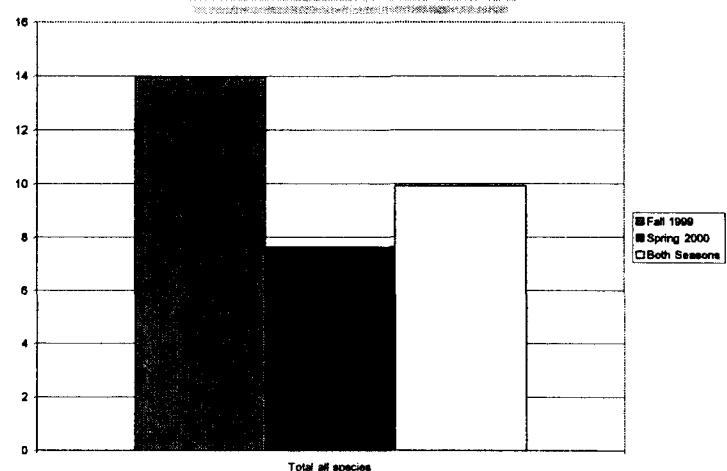
CPUE for Fish Collected by Gillnet from the PRSA



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CPUE for Fish Collected by Gillnet from the PRSA

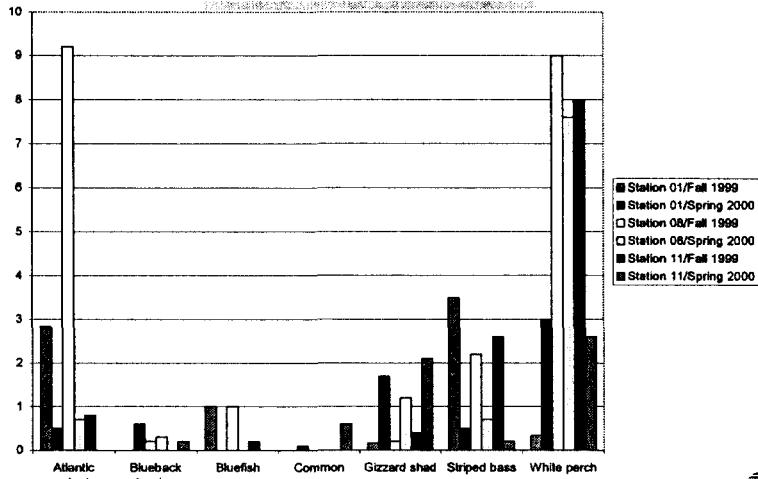


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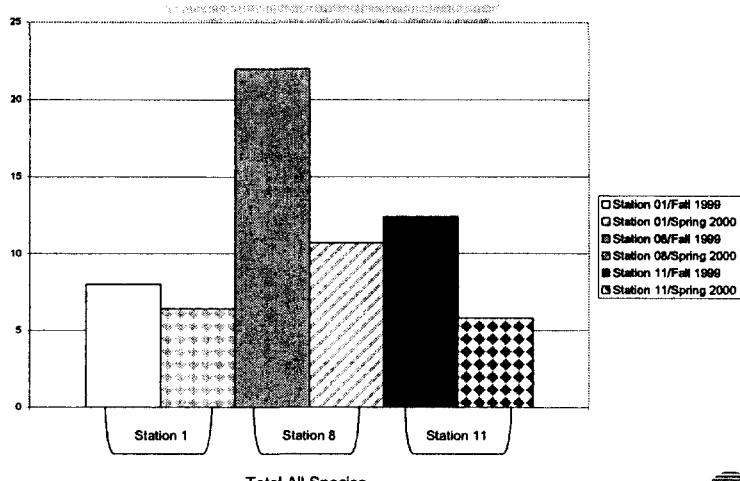
CPUE for Fish Collected by Gillnet from the PRSA by Station and Season



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CPUE for All Fish Species Collected by Gillnet from the PRSA by Station and Season



14

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Summary of Lengths and Weights for Fish Collected from the PRSA

	Combined Fall 1999 and Spring 2000						
	Length (mm)			Weight (g)			
	N*	Range	Mean	SD*	Range	Mean	SD*
American eel	20	230 - 630	366	102	20 - 499	120	119
Atlantic menhaden	57	86 - 375	307	67	9 - 691	340	154
Blueback herring	9	225 - 265	240	13	95 - 197	130	29
Bluefish	6	176 - 335	247	68	53 - 112	99.7	23
Brown bullhead	2	278 - 280	279	1.4	320 - 321	321	0.71
Channel catfish	1	--	193	--	--	78	--
Common carp	7	460 - 730	562	88	1,400 - 3,487	2573	717
Gizzard shad	50	352 - 495	442	29	391 - 1,763	1103	275
Striped bass	48	206 - 730	396	137	88 - 3,682	933	924
Weakfish	2	220 - 234	227	10	102 - 143	123	29
White catfish	4	122 - 360	280	109	237 - 764	482	244
White perch	164	132 - 310	206	40	41 - 428	161	90
White sucker	1	--	425	--	--	965	--

Notes:

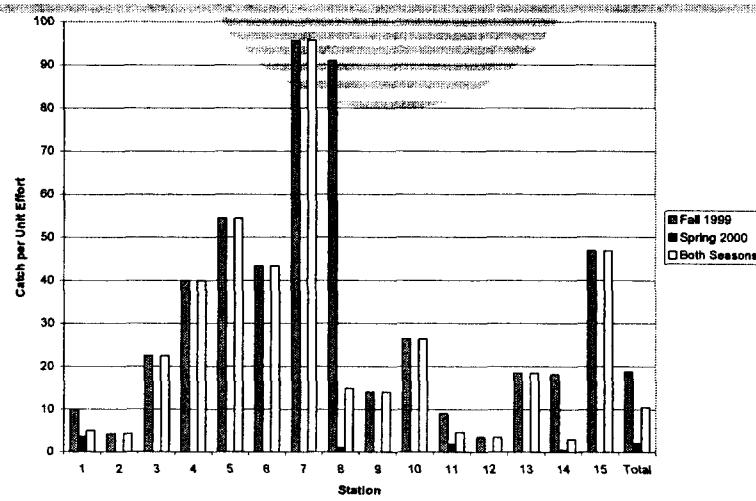
* Only intact fish for which complete measurements were available (length, weight) were included in this analysis.

SD = Standard Deviation

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CPUE for Mummichog Collected from the PRSA



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Summary of Length and Weight Data for Mummichog Collected from the PRSA

Station	Length Range		Length SD *	Weight Range		Average Weight (g)	Weight SD *
	(n)	(mm)		(mm)	(g)		
Fall 1999	1233	41 - 114	66.7	14.4	1.0 - 21	4.7	3.9
Spring 2000	158	45 - 110	69.8	14.4	1.0 - 19	4.7	3.5
Combined Total	1391	41 - 114	67.1	14.4	1.0 - 21	4.7	3.9

Station	Length Range		Length SD *	Weight Range		Average Weight (g)	Weight SD *
	(n)	(mm)		(mm)	(g)		
Fall 1999	1785	40 - 117	70.1	16.6	1.0 - 31	5.7	5.0
Spring 2000	157	45 - 130	79.2	16.5	1.0 - 28	7.9	5.0
Combined Total	1972	40 - 130	70.8	16.8	1.0 - 31	5.9	5.1

Station	Sex Ratio	
	M : F	
Fall 1999	1 : 1.45	
Spring 2000	1.0 : 0.99	
Combined Total	1 : 1.40	

Notes:

* SD - standard deviation



CPUE for Blue Crab Collected by Crab Trap from the PRSA

Sampling Event	No. of Crabs Collected ^a	No. of Traps Set	CPUE
Fall 1999	1269	262	4.84
Spring 2000	231	88	2.63
Combined Total	1500	350	4.29

Notes:

^aThis number includes crabs collected that were not measured in length-weight analysis.



Summary of Lengths and Weights for Blue Crab Collected from the PRSA

Sampling Event	No. of Crabs	Length Range (mm)	Length Average (mm)	Length SD^a	Weight Range (g)	Weight Average (g)	Weight SD^a
Fall 1999	1,210	64 - 192	121	16	34 - 269	106	39
Spring 2000	229	161 - 158	110	16	13 - 217	85	29
Combined Total	1,439	61 - 192	119	16	13 - 269	103	38

Notes:

^a SD = Standard Deviation

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Supplemental Fish Collection Program – August 2001

- Not a community survey
- Focused collection effort for supplemental fish tissue data – edible fillets for human health risk assessment
- Target species:
 - American eel
 - Catfish (i.e., catfish or bullhead)
 - Carp
- Multiple sampling gear types
- One week sampling effort

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Preliminary Sediment Quality Triad (SQT) Assessment



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The Sediment Quality Triad

Sediment Chemistry
(contaminant analyses)

Sediment Toxicity
(laboratory
bioassays)

Benthic
Invertebrate
Communities
(community analyses)

SQT Potential Scenarios

Contamination	Toxicity	Alteration	Scenario
+	+	+	Strong evidence for impacts from chemical contamination
-	-	-	Strong evidence for no impacts from chemical contamination
+	-	-	Chemical contaminants are not toxic or bioavailable
-	+	-	Unmeasured chemical or physical conditions exist that are causing toxicity
-	-	+	Impacts are not caused by chemical contamination
+	+	-	Chemical contaminants may be causing toxicity
-	+	+	Unmeasured chemical or physical conditions exist that are causing toxicity and community impacts
+	-	+	Chemical contaminants are not bioavailable or community alterations are not due to toxic chemicals

Notes:

"+" = contamination, toxicity, and/or community alterations present

"-" = contamination, toxicity, and/or community alterations absent

3



Objectives

- Compare sediment quality between the PRSA and Mullica River reference area
- Develop a qualitative, weight-of-evidence description for each PRSA station
- Rank and compare relative sediment quality among stations
- Identify which physico-chemical variables may influence sediment toxicity and/or benthic community alterations in PRSA

4



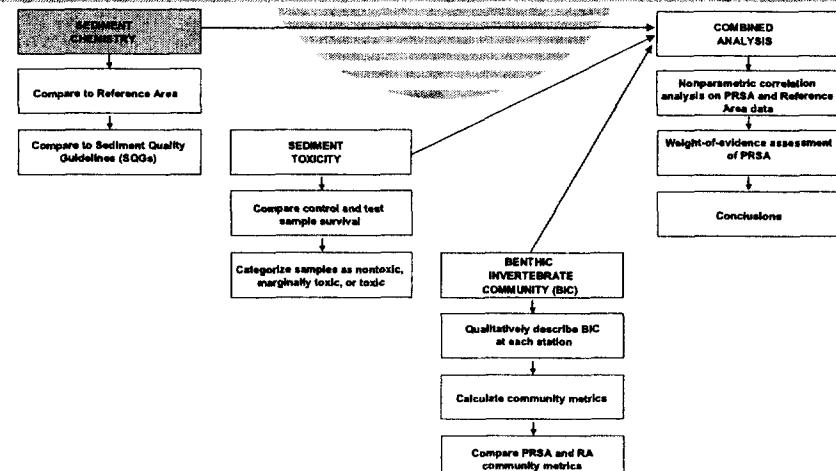
Methods

- Chemistry, toxicity, benthic community analyses documented in previous presentations
- Preliminary statistical analyses
 - Comparison of PRSA to Reference Area (RA)
 - Sediment quality guideline quotients (SQGQs)
 - Nonparametric Spearman correlations
- PRSA station classifications
- Weight-of-evidence assessment

5



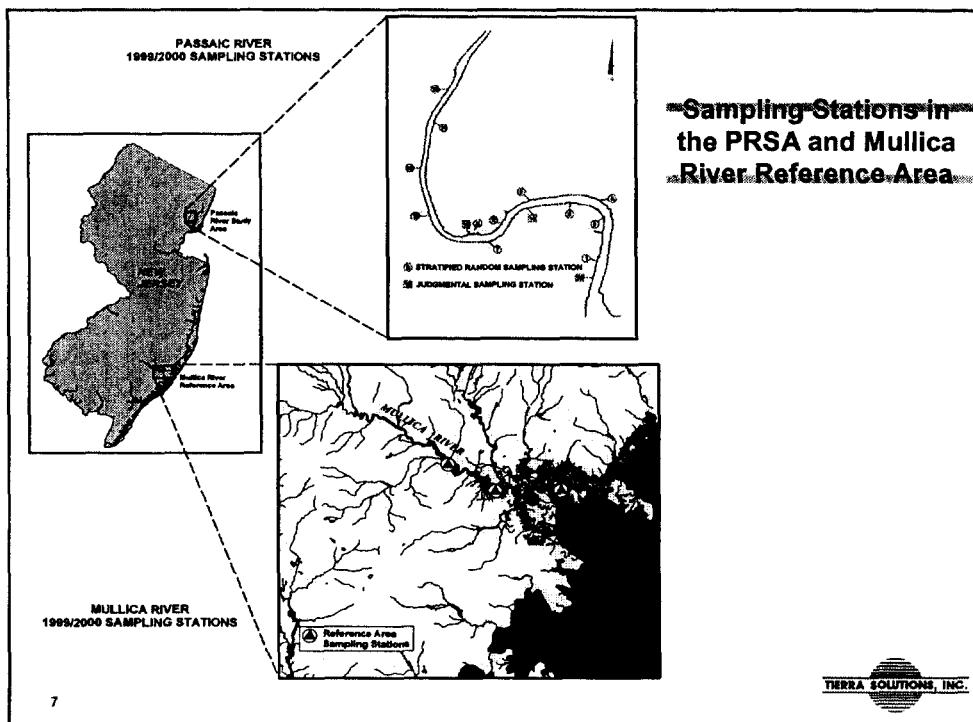
Steps in the SQT



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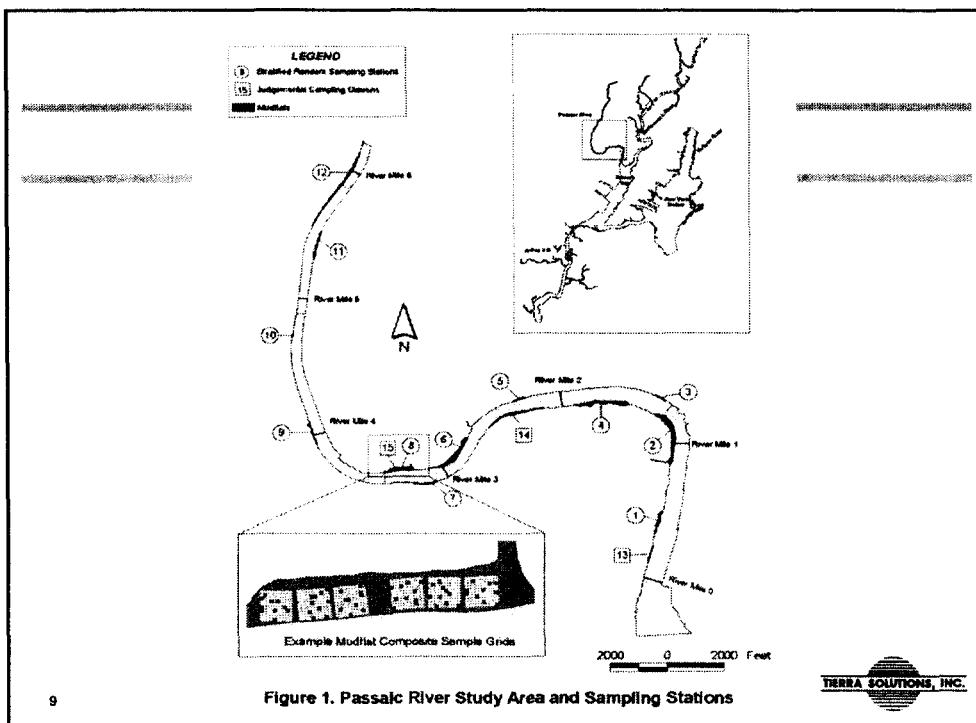
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PRSA Sediment Chemistry Data

- Described in detail in May 29, 2002 presentation
- Chemistry data from central sampling grid at each ESP station used in SQT – synoptically collected with toxicity and benthic invertebrate community data

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Chemicals Evaluated in the SQT

Inorganic Chemicals	Miscellaneous	Polyaromatic Hydrocarbons (PAHs)
Aluminum	Ammena Nitrogen	High Molecular Weight PAHs (13) ^a [H-PAHs]
Antimony	Percent Fines	Low Molecular Weight PAHs (13) ^a [L-PAHs]
Arsenic	Total Organic Carbon	Total PAHs (13) ^a
Barium	Salinity	
Beryllium	pH	
Cadmium	Organotins	Semivolatile Compounds
Calcium	Dibutyltin	1,4-Dichlorobenzene
Chromium	Monobutyltin	2,4-Dichlorophenol
Cobalt	Tributyltin	bis(2-Ethylhexyl)phthalate
Copper		Butyl benzyl phthalate
Iron		Carbazole
Lead	Pesticides/Herbicides	Dibenzofuran
Magnesium	Total DDT	Dibenzothiophene
Manganese		Di-n-Butylphthalate
Mercury	Polychlorinated Biphenyls (PCBs)	Di-n-Octylphthalate
	Total PCBs - Sum of Homologue Groups	N-Nitrosodiphenylamine
Nickel		
Potassium		
Selenium	Polychlorinated Dibenzo-p-Dioxins and Furans (PCDD/Fs)	
Silver	WHO TEQ(Fish)	
Sodium		
Thallium		
Vanadium		
Zinc		

Notes:
^a Calculated using a limited congener set (13 PAHs) as described in Long et al., 1995.

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Sediment Quality Guideline Quotients (SQGQ)

SQGQ	
ER-M Quotient (PAH categories)	Calculated using ER-Ms for the following chemicals: As, Cd, Cr, Cu, Pb, Hg, H-PAH, L-PAH, Total PAHs, Total PCBs (homologue groups), Ag, Total DDT, and Zn. PAH categories calculated using the method of Long et al. (1995) with only 13 PAHs as listed in the individual PAHs ER-MQ below.
ER-M Quotient (PAH individual)	Calculated using ER-Ms for the following chemicals: As, Cd, Cr, Cu, Pb, Hg, 2-Methylnaphthalene, Dibenz(a,h)naphthalene, Acenaphthene, Acenaphthylene, Anthracene, Benz[a]anthracene, Benzo[a]pyrene, Chrysene, Fluoranthene, Fluorene, Naphthalene, Phenanthrene, Pyrene, Total PCBs (homologue groups), Ag, Total DDT, and Zn.
SQGQ ER-M + Mn	Calculated using ER-Ms for the following chemicals: As, Cd, Cr, Cu, Pb, Hg, H-PAH, L-PAH, Total PAHs, Total PCBs (homologue groups), Ag, Total DDT, and Zn plus a benchmark value for Mn. PAH categories calculated using the method of Long et al. (1995) with only 13 PAHs as listed in the individual PAHs ER-MQ above.
SQGQ ER-M + BEP	Calculated using ER-Ms for the following chemicals: As, Cd, Cr, Cu, Pb, Hg, H-PAH, L-PAH, Total PAHs, Total PCBs (homologue groups), Ag, Total DDT, and Zn plus a benchmark value for bis(2-ethylhexyl)phthalate. PAH categories calculated using the method of Long et al. (1995) with only 13 PAHs as listed in the individual PAHs ER-MQ above.
SQGQ ER-M + PCDD/F TEQ	Calculated using ER-Ms for the following chemicals: As, Cd, Cr, Cu, Pb, Hg, H-PAH, L-PAH, Total PAHs, Total PCBs (homologue groups), Ag, Total DDT, and Zn plus a benchmark value for PCDD/F TEQ. PAH categories calculated using the method of Long et al. (1995) with only 13 PAHs as listed in the individual PAHs ER-MQ above.
SQGQ All benchmarks	Calculated using ER-Ms for the following chemicals: As, Cd, Cr, Cu, Pb, Hg, H-PAH, L-PAH, Total PAHs, Total PCBs (homologue groups), Ag, Total DDT, and Zn plus benchmark values for Mn, and PCDD/F TEQ. PAH categories calculated using the method of Long et al. (1995) with only 13 PAHs as listed in the individual PAHs ER-MQ above.

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SQGQs for the PRSA and RA

Station	ER-M Quotient (PAH categories)	ER-M Quotient (PAH individual)	SQGQ ER-M + Mn	SQGQ ER-M + BEP	SQGQ ER-M + PCDD/F TEQ	SQGQ All Benchmarks
PRSA						
1	1.9	1.5	1.8	2.2	1.8	1.9
2	1.6	1.4	1.5	1.6	1.5	1.5
3	0.86	0.79	0.81	1.0	0.80	0.86
4	1.9	1.6	1.8	2.2	1.8	2.0
5	1.7	1.6	1.6	1.8	1.5	1.6
6	1.8	1.6	1.7	1.9	1.7	1.7
7	2.5	1.9	2.3	2.7	2.3	2.4
8	1.8	1.5	1.7	2.0	1.7	1.8
9	3.3	2.5	3.1	3.4	3.1	3.0
10	1.9	1.5	1.8	2.1	1.8	1.9
11	2.5	2.2	2.3	3.1	2.3	2.8
12	2.0	1.8	1.9	2.3	1.9	2.0
13	2.1	1.8	1.9	2.3	1.9	2.1
14	2.0	1.6	1.9	2.1	1.8	1.9
15	2.1	1.8	2.0	2.3	2.0	2.1
Reference Area						
21	0.22	0.15	0.22	0.23	0.21	0.21
22	0.15	0.10	0.15	0.16	0.14	0.15
23	0.19	0.12	0.19	0.19	0.17	0.18

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Classification System for PRSA and Reference Area Sediments based on SQGs/SQGQs

Sediment Type	Number of SQGs Exceeded	Average SQGQ Value
1	0	≤ 0.50
2	1-4	0.51-1.0
3	5-9	1.1-2.4
4	≥ 10	≥ 2.41

Notes:

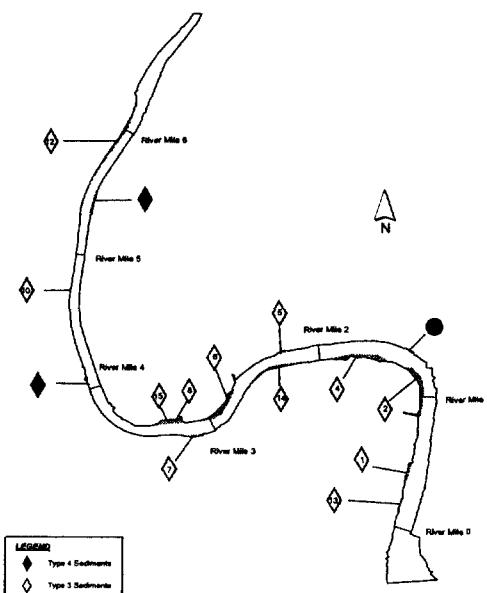
PRSA-specific classification system.

Reference Area stations contain Type 1 sediments.

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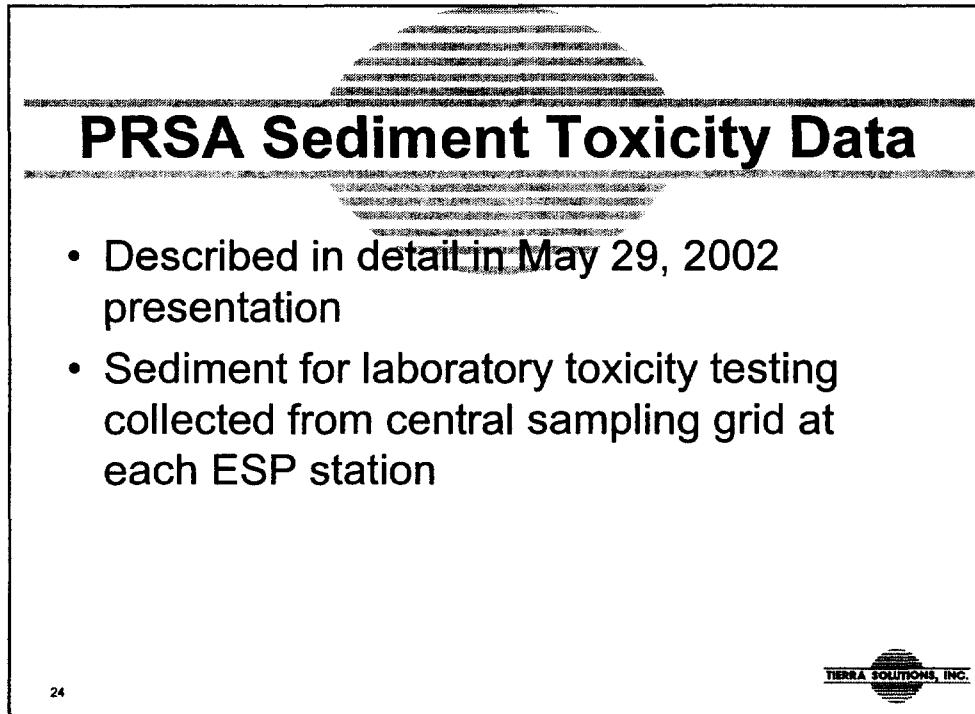
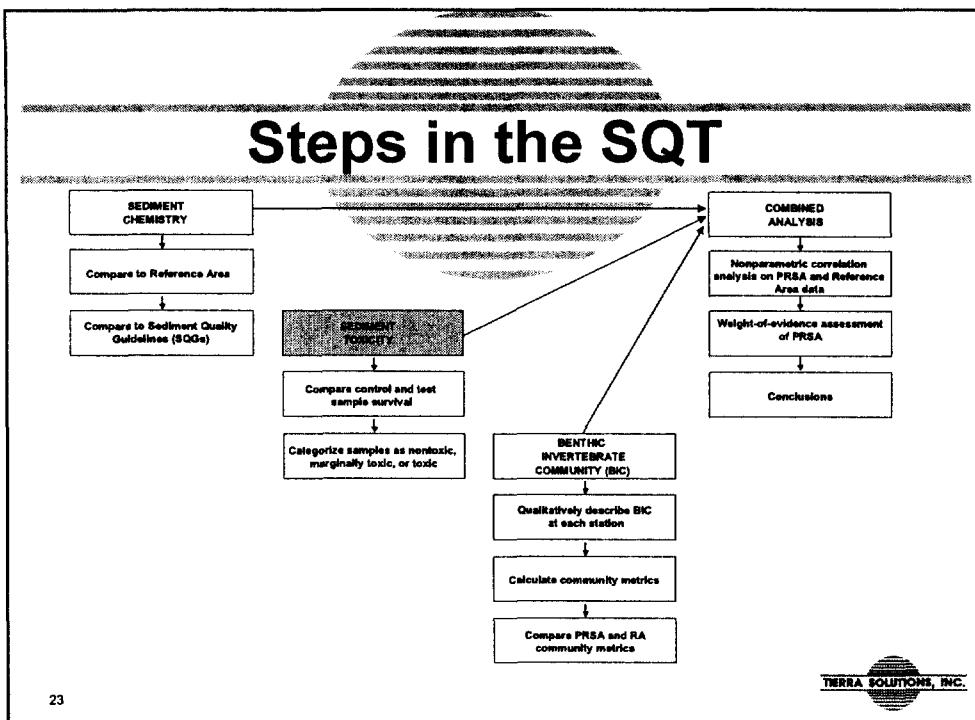
SQGQ Classification in the PRSA



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Sediment Sample Toxicity

A sample is considered:

- Nontoxic if mean survival was not significantly different ($p > 0.05$) from negative controls
- Marginally toxic if mean survival was significantly lower than in negative controls ($p < 0.05$) but exceeded 80% of average survival in controls (amphipods) or exceeded 64% of average survival in controls (polychaetes)
- Highly toxic if mean survival was significantly lower than in negative controls ($p < 0.05$) and < 80% of average survival in controls (amphipods) or < 64% of average survival in controls (polychaetes)

Source: Long et al., 2000

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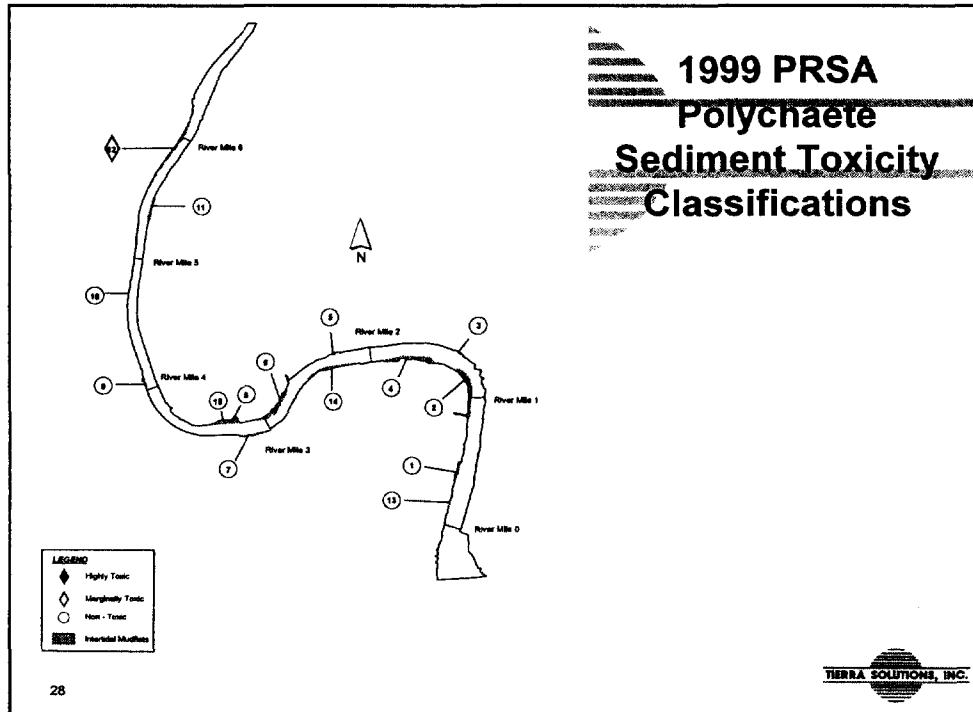
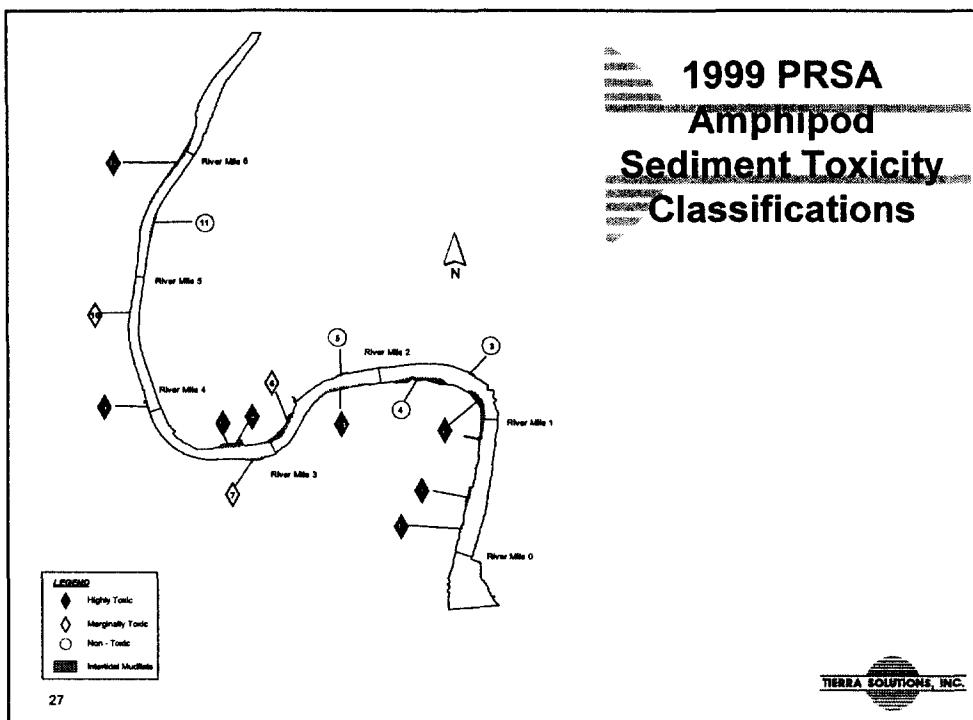
1999 PRSA Sediment Toxicity Testing Results

Station	Average Percent Survival (Amphipod)	Toxicity		Toxicity Category (Polychaete Survival)
		Category (Amphipod Survival)	Average Percentage (Polychaete Survival)	
PRSA				
1	70	Highly toxic	100	Nontoxic
2	68	Highly toxic	100	Nontoxic
3	83	Nontoxic	100	Nontoxic
4	85	Nontoxic	100	Nontoxic
5	79	Nontoxic	96	Nontoxic
6	72	Marginally toxic	100	Nontoxic
7	75	Marginally toxic	96	Nontoxic
8	43	Highly toxic	100	Nontoxic
9	46	Highly toxic	92	Nontoxic
10	75	Marginally toxic	100	Nontoxic
11	78	Nontoxic	92	Nontoxic
12	46	Highly toxic	84	Marginally Toxic
13	70	Highly toxic	82	Nontoxic
14	46	Highly toxic	100	Nontoxic
15	68	Highly toxic	96	Nontoxic
Reference Area				
21	95	Nontoxic	100	Nontoxic
22	92	Nontoxic	100	Nontoxic
23	92	Nontoxic	96	Nontoxic
Laboratory Controls				
1	89		100	
2	89		96	

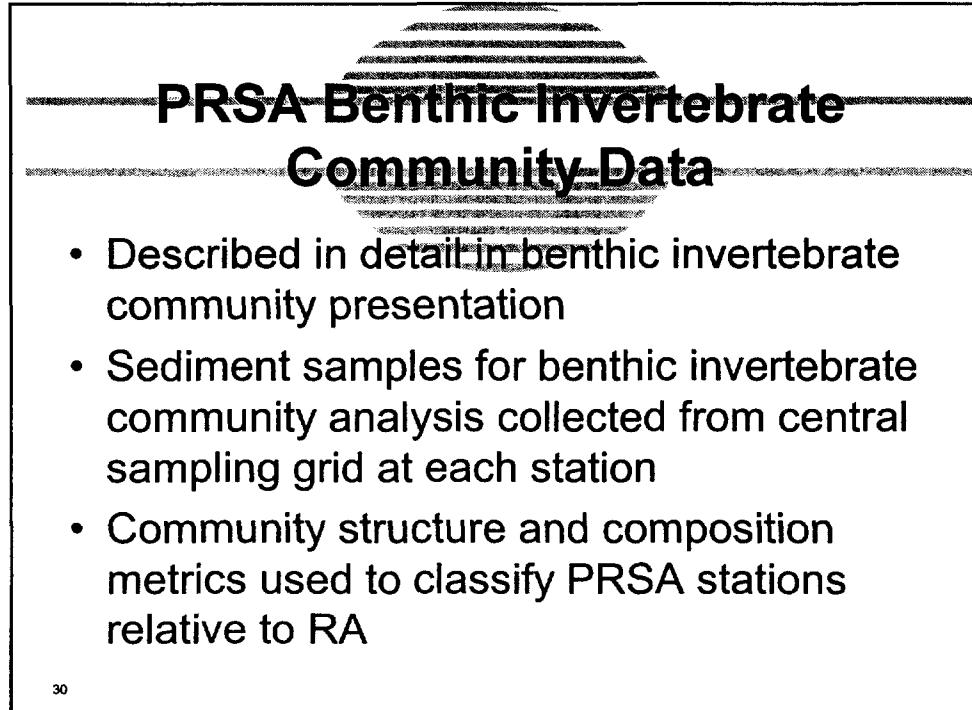
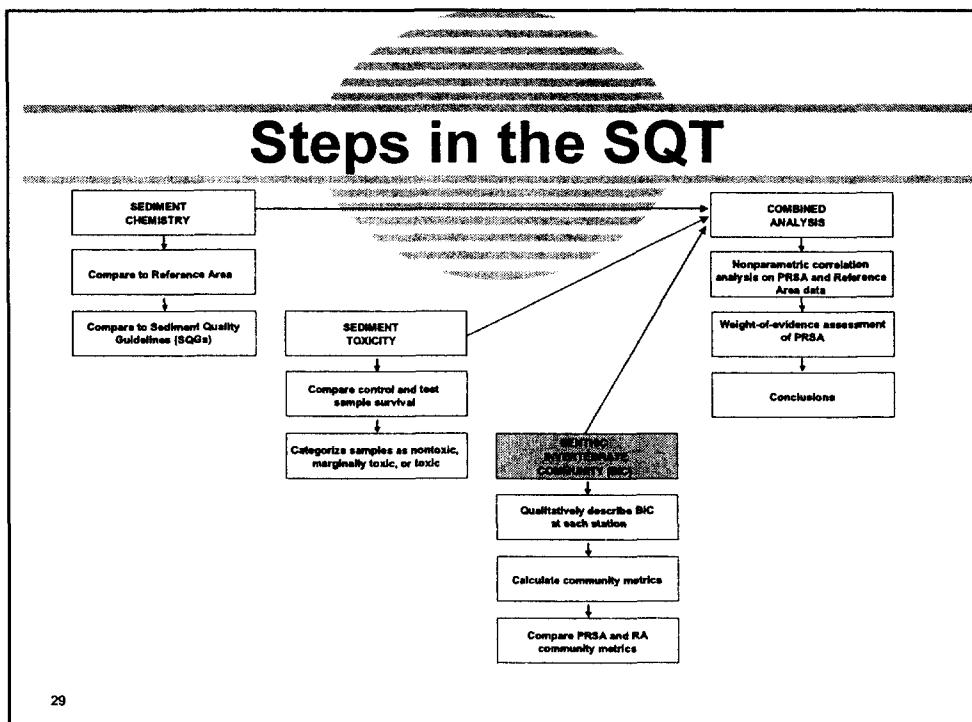
Note:
Data was arcsine square root transformed, which made the data meet ANOVA assumptions than a one-tail t-test with equal variance was performed to see which stations were significantly different from the negative controls.

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Qualitative Ranks for Each PRSA Station Compared to Reference Area

Metric	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15
No. of Individuals ¹	poor	good	poor												
No. of Taxa	good														
Abundance of Crustaceans	poor														
Abundance of Tolerant Taxa ¹	poor	good	poor												
Pielou's Evenness	poor	good	poor												
Shannon's H'	poor	poor	good	poor											
Virginia H'	poor	poor	good	poor											
Bilobian H'	poor	poor	good	poor											
Swarz Dominance Index	good														

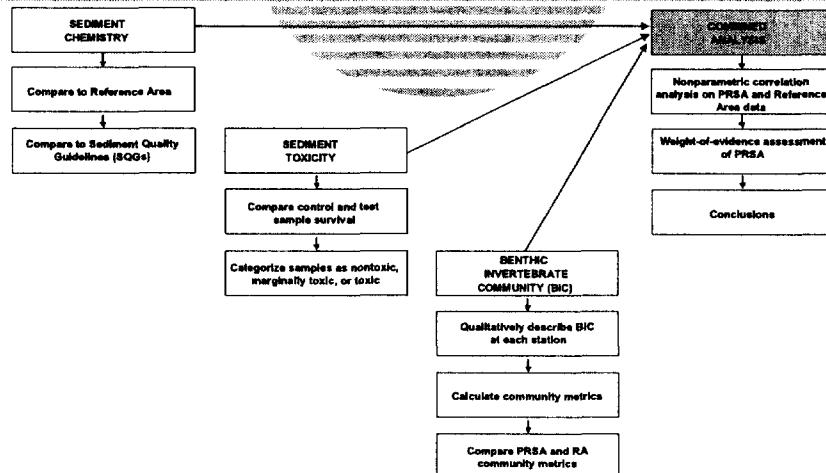
Note:

¹ For the number of individuals and abundance of tolerant taxa metrics, the following ranks were assigned to each PRSA and Reference Area comparison: a) above reference range = poor; b) within reference range = good; c) below reference range = excellent. For the remaining metrics, the following ranks were assigned for each PRSA/Reference Area comparison: a) above reference area = excellent; b) within reference area = good; c) below reference area = poor.

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Steps in the SQT



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Spearman Rank Correlations of Sediment Quality Guidelines and Toxicity and Benthic Invertebrate Community Parameters (n=18)

	Amphipod Survival	Number of Individuals	Number of Species	Wetland Diversity	Pearson's Product-Moment Correlation Coefficient (r)	Spearman's Rank Correlation Coefficient (rho)	Virginia Province Index	Percent Crustacea	Percent Pollution-Tolerant Organisms
ER-MO (PAH categories)	-0.535	0.235	-0.374	-0.322	0.062	-0.316	-0.047	0.000	0.42
ER-MO (PAH individual)	-0.491	0.239	-0.311	-0.369	0.068	-0.263	-0.061	2.533	0.52
SOGO ER-M + Mn	-0.565	0.275	-0.406	-0.361	0.060	-0.355	-0.060	0.000	0.42
SOGO ER-M + BEP	-0.523	0.282	-0.372	-0.341	0.011	-0.334	-0.061	0.000	0.42
SOGO ER-M + PCDD/F TEQ	-0.544	0.261	-0.364	-0.350	0.024	-0.345	-0.065	0.000	0.42
SOGO All benchmarks	-0.497	0.236	-0.366	-0.326	0.031	-0.321	-0.037	0.000	0.42

Notes:

Using a Bonferroni adjusted alpha level based on the number of analytes (8), p must be ≤ 0.01 for a significant correlation to exist.

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Concordance of Triad Components

Station	Sediment Type	Sediment Toxicity	BIC Condition	Component Agreement
13	3	Highly toxic	Good - Excellent	No
1	3	Highly toxic	Poor	Yes
2	3	Highly toxic	Poor - Good	Yes
3	2	Nontoxic	Good	Yes
4	3	Nontoxic	Poor - Good	No
14	3	Highly toxic	Poor	Yes
5	3	Nontoxic	Good	No
6	3	Marginally toxic	Good	No
7	3	Marginally toxic	Poor - Good	No
8	3	Highly toxic	Good	No
15	3	Highly toxic	Poor	Yes
9	4	Highly toxic	Poor - Good	Yes
10	3	Marginally toxic	Poor	Yes
11	4	Nontoxic	Poor	No
12	3	Highly toxic	Poor	Yes

Note:

Stations ordered from downstream to upstream in PRSA.

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SQT Uncertainties

- Unidentified/unanalyzed chemicals could be impacting sediment toxicity and benthic invertebrate community structure
- Seasonal effects on sediment toxicity, sediment chemistry, and benthic invertebrate community structure and composition
- Role of chemical synergy in sediment toxicity and benthic invertebrate community structuring
- No SQGs available for many detected chemicals

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Overall Weight-of-Evidence Conclusions

- Elevated levels of chemicals found at many PRSA stations relative to Reference Area
- No clear spatial gradients in chemical concentrations present in the PRSA
- Sediment quality guidelines exceeded for a number of chemicals at multiple stations
- Amphipod toxicity detected in PRSA samples – no clear spatial gradient
- Amphipod toxicity not likely caused by single chemical or physical factor
- PRSA benthic invertebrate community structure and composition generally “poor” relative to the Reference Area

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Next Steps

- Multi-variate statistical analyses
- Evaluate SQT with respect to TIE results



Phase I Toxicity Identification Evaluation (TIE)

Preliminary Assessment



Objectives

- Determine if one or more chemical classes appear responsible for sediment toxicity to benthic invertebrates in the PRSA
- Perform an investigation to supplement the sediment quality triad (SQT) assessment being performed under the CERLCA RI/FS

Field Sampling Methods

- July 2000 sampling event
- Five stations in the PRSA — corresponding to ESP stations 7, 11, 12, 13, and 14
- Stations selected for apparent differences in predominant chemical contaminant mixtures
- Surface sediment samples collected

3



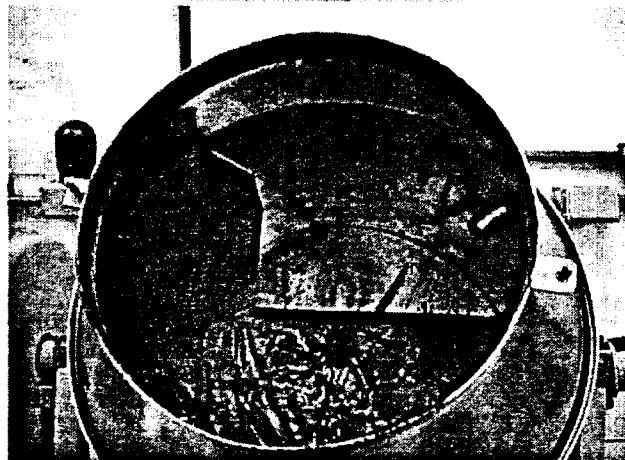
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Locations of TIE Sample Stations



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Customized Stainless Steel Mixer Used to Homogenize PRSA Sediment Samples



5

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Laboratory Methods

- Followed USEPA (1996) Phase I Marine TIE procedures – pore water manipulations
- Contaminant chemistry analyses (comparable to CERCLA RI/FS) performed on sediment and pore water samples
- Sediment and pore water toxicity tests using the amphipod *Ampelisca abdita*
- Additional Microtox® pore water toxicity tests

6

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TIE Methods Summary

- Initial and baseline toxicity tests
- Multiple pore water manipulations – results compared to baseline
- Five pore water dilutions for each manipulation (0, 25%, 50%, 75%, 100%)
- Dose-response relationships examined – LC50s calculated



Summary of Phase I TIE Manipulations Performed on Pore Water Samples from Each Station

Manipulation Type	Chemical Focus of Manipulation
Filtration	To remove toxicity associated with particulate-bound toxicants
Aeration	To remove toxicity associated with volatile organic compounds, sulfides, and ammonia
Ethylenediaminetetraacetic acid (EDTA) chelation	To remove toxicity associated with metals
Sodium thiosulfate ($\text{Na}_2\text{S}_2\text{O}_3$) treatment	To remove toxicity associated with oxidants (i.e., chlorine), and some metals
Solid-phase extraction (SPE) through a C-18 column/follow-up elution	To remove toxicity associated with non-polar organic compounds such as pesticides, PCDD/Fs, and PAHs
Graduated pH adjustment to pH 7, pH 8, and pH 9	To remove pH-dependent toxicants such as ammonia and hydrogen sulfide
SPE through a cation exchange resin/follow-up elution	To remove toxicity associated with divalent metals
<i>Ulva lactuca</i> treatment	To remove toxicity associated primarily with ammonia, with some secondary removal of hydrogen sulfide and organic compounds



Results

- Percent amphipod survival in sediments was zero or near zero in each sample
- Pore water toxicity to amphipods varied between stations in initial and baseline tests
- No toxicity observed during baseline toxicity test (pore water) at Station 11
- Some post-manipulation toxicity tests had either high control mortality or no dose-response relationship

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Comparison of Initial and Baseline Study Results Using *Ampelisca abdita*

Station	Whole Sediment		Pore Water (48 hours LC50)	
	10 days (% survival)	Initial	Initial	Baseline*
7	0	24	29	
11	0	83	>100	
12	0	73	<13	
13	3	14	33	
14	0	35	75	

Notes:

* Baseline tests conducted in conjunction with TIE manipulation samples (48 hours after initial tests).

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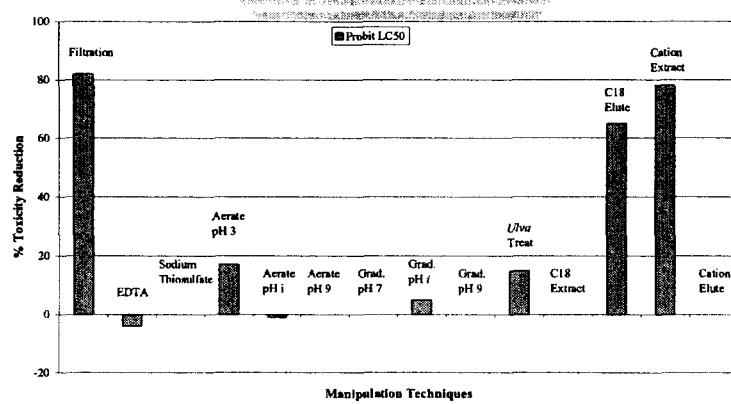
Results

- No toxicity in Microtox® tests in pore water samples from Stations 11, 13, and 14 – low toxicity at stations 7 and 12
- Suggests that the following are not likely toxicants:
 - Oxidants
 - Dissolved phase metals
 - Dissolved phase neutral organics

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Comparison of 48-Hour Toxicity Study Results for Phase I TIE Manipulations for Station 7 Based on LC50 Analysis



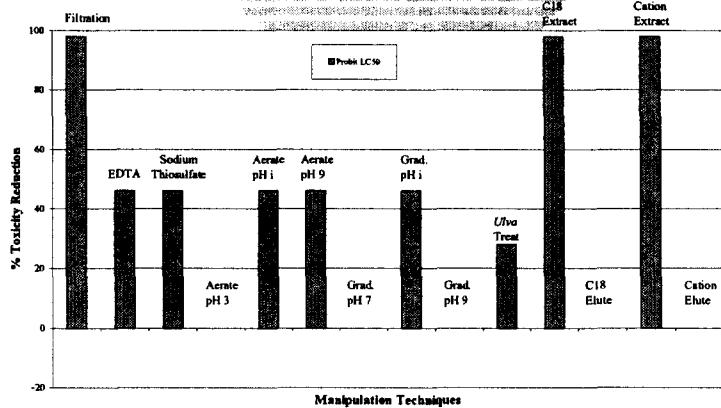
12

Manipulated fraction results compared to baseline results.

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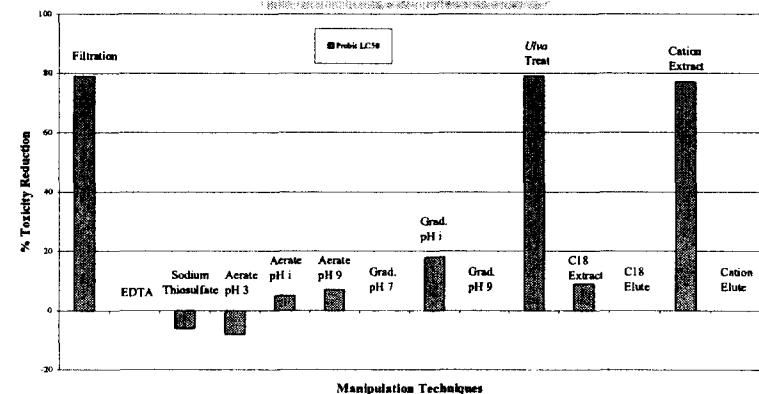
Comparison of 48-Hour Toxicity Study Results for Phase I TIE Manipulations for Station 12 Based on LC50 Analysis



Manipulated fraction results compared to baseline results.



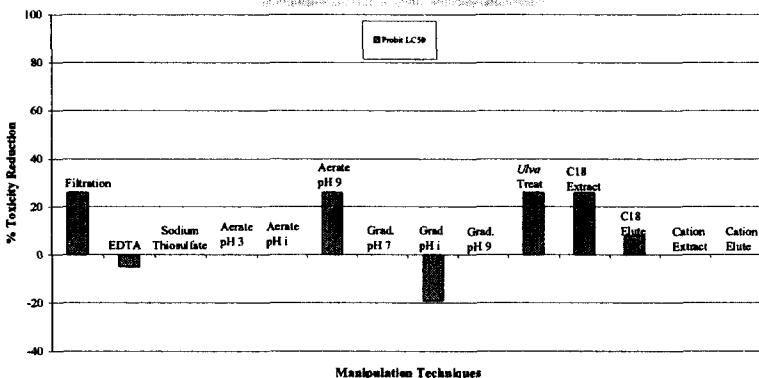
Comparison of 48-Hour Toxicity Study Results for Phase I TIE Manipulations for Station 13 Based on LC50 Analysis



Manipulated fraction results compared to baseline results.



Comparison of 48-Hour Toxicity Study Results for Phase I TIE Manipulations for Station 14 Based on LC50 Analysis



Manipulated fraction results compared to baseline results.

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TIE Results – Preliminary Interpretation

Manipulation	Key Chemical Class	Results		
		Station 11	Station 12	Station 13
Filtration	Particulates	++	++	+
Aeration	VOCs/Ammonia/Sulfides	+	+	+
EDTA Chelation	Metals	-	+	NR
Na ₂ S ₂ O ₃ Treatment	Metals	NR	+	-
pH Adjustments	Ammonia/Sulfides	-	+	+/-
C-18 SPE	Nonpolar Organic Compounds	NR	++	+
Cation Exchange SPE	Metals	++	++	++
Uva /aerace Treatment	Ammonia/Sulfides	+	+	++

TIE Interpretation Regarding Possible Causes of Toxicity:

1.	Particle-associated toxicity	Particle-associated nonpolar organic compounds	Particle-associated nonpolar organic compounds	Particle-associated nonpolar organic compounds
2.	Particle-associated metals	Particle-associated metals	Ammonia	Ammonia
3.	Ammonia	Ammonia	Low response = other contributors	Low response = other contributors

Notes:

- ++ = indicates strong toxicity reduction.
- + = indicates low to moderate toxicity reduction.
- = indicates ineffective toxicity reduction.
- NR = no dose-response relationship or high control mortality occurred in this manipulation.

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Preliminary Conclusions

- Toxicity characteristics at stations exhibiting baseline toxicity were consistent with particle-associated chemicals
 - Toxicity removed primarily by filtration
 - Sediment tests had higher toxicity than pore water tests
 - Microtox® toxicity low or zero
- Ammonia may be a seasonal contributor to toxicity

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Next Steps

- Evaluate TIE results with respect to sediment and pore water chemistry analyses (e.g., toxic units assessment)
- Final interpretation
- Integration with SQT

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Wrap-Up Discussion

Action Items and Assignments



Data Gaps for PRRI

- Water chemistry data
- CSO data
- Tissue chemistry data (PRSA to Dundee Dam)
- Sediment chemistry data (PRSA to Dundee Dam)
- Sediment toxicity data (PRSA to Dundee Dam)
- Quantitative habitat/bird relationship for restoration
- Habitat characterization (PRSA to Dundee Dam)
- Geotechnical/hydrodynamic field data (PRSA to Dundee Dam)
- Other



Next Meeting

- Potential meeting date: Friday, November 8, Silver Spring, MD
- Dioxin sources identification analyses
- Technical Work Group(s) establishment
- Trustee presentation/discussion
- Other?

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ATTENDANCE LIST
PRSA Technical Meeting
September 26, 2002
NJTPA

Name	Organization	Telephone
Dave Ludwig	BBL Sciences	410.295.1205
Rick Winfield	USEPA	212.637.4362
Carol Jenkins	VET for America	713 758 2528
Dennis Farley	ElgarP	908 901-0112

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ATTENDANCE LIST
 PRSA Technical Meeting
 September 26, 2002
 NJTPA

Name	Organization	Telephone
GENE MANCINI	TSI consultant	805 987 7152
RICK McNUTT	Tierra Solutions	732-246-5849
Cliff Firstenberg	Tierra Solutions	757-258-7720
Joe Steinbacher	NOAA/DAC	301 713 3038 x182
LISA Rosman	NOAA/CPRD	212 637 3259
Tom Rutherford	NOAA/DAC	201-713-3038 x186
Sharm Jaffess	USEPA	212-637-4396
B Danjue	DOS	212-514-5413
BEN TROTTER	NJDEP ONRR	609 984 6155
ROB RICKER	NOAA/DAC	301-713-3038 x 131
Karen Rudd	EPA/OPC	212-637-3106
Roselle Henn	USACE	
Sandra Brown	USFWS	609-646-9310 x33
LISA BARON	OMR/NJDOE	609-530-4779
CHRISTIAN STITT	DESA -USEPA	732-321-6676
Tim Iannuzzi	BBL Sciences	410-295-1205
Tim Kubick	USFWS	609 646 9310 x26
SANDY CABBE	Tierra Solutions, Inc.	732-246-5848
Joan Daufeld	NJ Div. of Law	609 984 4654

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